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(71) Applicant:
F. HOFFMANN-LA ROCHE AG
4070 Basel (CH)

(72) Inventors:
• **Hoshino, Tatsuo**
Kamakura-shi, Kanagawa-ken 248-0027 (JP)
• **Ojlma, Kazuyuki**
Fujisawa-shi, Kanagawa-ken 251-0877 (JP)
• **Setoguchi, Yutaka**
Fujisawa-shi, Kanagawa-ken 251-0033 (JP)

(74) Representative: **Braun, Axel et al**
F.Hoffmann-La Roche AG
Patent Department (PLP),
124 Grenzacherstrasse
4070 Basel (CH)

(54) **Dna sequences encoding enzymes involved in production of isoprenoids**

(57) The present invention is directed to an isolated DNA sequence coding for an enzyme involved in the mevalonate pathway or the pathway from isopentenyl pyrophosphate to farnesyl pyrophosphate, vectors or plasmids comprising such DNA, hosts transformed by either such DNAs or vectors or plasmids and a process for the production of isoprenoids and carotenoids by using such transformed host cells.

Description

[0001] The present invention relates to molecular biology for the manufacture of isoprenoids and biological materials useful therefor.

[0002] Astaxanthin is known to distribute in a wide variety of organisms such as animal (e.g. birds such as flamingo and scarlet ibis, and fish such as rainbow trout and salmon), algae and microorganisms. It is also recognized that astaxanthin has a strong antioxidation property against oxygen radical, which is expected to apply to pharmaceutical usage to protect living cells against some diseases such as a cancer. Moreover, from a viewpoint of industrial application, a demand for astaxanthin as a coloring reagent is increasing especially in the industry of farmed fish, such as salmon, because astaxanthin imparts distinctive orange-red coloration to the animals and contributes to consumer appeal in the marketplace.

[0003] *Phaffia rhodozyma* is known as a carotenogenic yeast strain which produces astaxanthin specifically. Different from the other carotenogenic yeast, *Rhodotorula* species, *Phaffia rhodozyma* (*P. rhodozyma*) can ferment some sugars such as D-glucose. This is an important feature from a viewpoint of industrial application. In a recent taxonomic study, a sexual cycle of *P. rhodozyma* was revealed and its teleomorphic state was designated under the name of *Xanthophyllomyces dendrorhous* (W.I. Golubev; Yeast 11, 101 - 110, 1995). Some strain improvement studies to obtain hyper producers of astaxanthin from *P. rhodozyma* have been conducted, but such efforts have been restricted to employ the method of conventional mutagenesis and protoplast fusion in this decade. Recently, Wery *et al.* developed a host vector system using *P. rhodozyma* in which a non-replicable plasmid was used to be integrated onto the genome of *P. rhodozyma* at the locus of ribosomal DNA in multicopies (Wery *et al.*, Gene, 184, 89-97, 1997). And Verdoes *et al.* reported more improved vectors to obtain a transformant of *P. rhodozyma* as well as its three carotenogenic genes which code the enzymes that catalyzes the reactions from geranylgeranyl pyrophosphate to β -carotene (International patent WO97/23633). The importance of genetic engineering method on the strain improvement study of *P. rhodozyma* will increase in near future to break through the reached productivity by the conventional methods.

[0004] It is reported that the carotenogenic pathway from a general metabolite, acetyl-CoA consists of multiple enzymatic steps in carotenogenic eukaryotes as shown in Fig.1. Two molecules of acetyl-CoA are condensed to yield acetoacetyl-CoA which is converted to 3-hydroxy-3-methylglutaryl-CoA (HMG-CoA) by the action of 3-hydroxymethyl-3-glutaryl-CoA synthase. Next, 3-hydroxy-3-methylglutaryl-CoA reductase converts HMG-CoA to mevalonate, to which two molecules of phosphate residues are then added by the action of two kinases (mevalonate kinase and phosphomevalonate kinase). Mevalonate pyrophosphate is then decarboxylated by the action of mevalonate pyrophosphate decarboxylase to yield isopentenyl pyrophosphate (IPP) which becomes a building unit of wide varieties of isoprene molecules which is necessary in living organisms. This pathway is called as mevalonate pathway taken from its important intermediate, mevalonate. IPP is isomerized to dimethylaryl pyrophosphate (DMAPP) by the action of IPP isomerase. Then, IPP and DMAPP converted to C₁₀ unit, geranyl pyrophosphate (GPP) by the head to tail condensation. In a similar condensation reaction between GPP and IPP, GPP is converted to C₁₅ unit, farnesyl pyrophosphate (FPP) which is an important substrate of cholesterol in animal and ergosterol in yeast, and of farnesylation of regulation protein such as RAS protein. In general, the biosynthesis of GPP and FPP from IPP and DMAPP are catalyzed by one enzyme called FPP synthase (Laskovics *et al.*, Biochemistry, 20, 1893-1901, 1981). On the other hand, in prokaryotes such as eubacteria, isopentenyl pyrophosphate was synthesized in a different pathway via 1-deoxyxylulose-5-phosphate from pyruvate which is absent in yeast and animal (Rohmer *et al.*, Biochem. J., 295, 517-524, 1993). In exclusive studies of cholesterol biosynthesis, it was shown that rate-limiting steps of cholesterol metabolism were in the steps of this mevalonate pathway, especially in its early steps catalyzed by HMG-CoA synthase and HMG-CoA reductase. The inventors paid their attention to the fact that the biosynthetic pathways of cholesterol and carotenoid share their intermediate pathway from acetyl-CoA to FPP, and tried to improve the rate-limiting steps in the carotenogenic pathway which might exist in the steps of mevalonate pathway, especially in early mevalonate pathway such as the steps catalyzed by HMG-CoA synthase and HMG-CoA reductase so as to improve the productivity of carotenoids, especially astaxanthin.

[0005] This invention is created based on the above endeavor of the inventors. In accordance with this invention, the genes and the enzymes involved in the mevalonate pathway from acetyl-CoA to FPP which are biological materials useful in the improvement of the astaxanthin production process are provided. This invention involves cloning and determination of the genes which code for HMG-CoA synthase, HMG-CoA reductase, mevalonate kinase, mevalonate pyrophosphate decarboxylase and FPP synthase. This invention also involves the enzymatic characterization as a result of the expression of such genes in suitable host organisms such as *E. coli*. These genes may be amplified in a suitable host, such as *P. rhodozyma* and their effects on the carotenogenesis can be confirmed by the cultivation of such a transformant in an appropriate medium under an appropriate cultivation condition.

[0006] According to the present invention, there are provided an isolated DNA sequence coding for an enzyme involved in the mevalonate pathway or the reaction pathway from isopentenyl pyrophosphate to farnesyl pyrophosphate. More specifically, the said enzyme are those having an activity selected from the group consisting of 3-hydroxy-

3-methylglutaryl-CoA synthase activity, 3-hydroxy-3-methylglutaryl-CoA reductase activity, mevalonate kinase activity, mevalonate pyrophosphate decarboxylase activity and farnesyl pyrophosphate synthase.

[0007] The said isolated DNA sequence may be more specifically characterized in that (a) it codes for the said enzyme having an amino acid sequence selected from the group consisting of those described in SEQ ID NOs: 6, 7, 8, 9 and 10, or (b) it codes for a variant of the said enzyme selected from (i) an allelic variant, and (ii) an enzyme having one or more amino acid addition, insertion, deletion and/or substitution and having the stated enzyme activity. Particularly specified isolated DNA sequence mentioned above may be that which can be derived from a gene of *Phaffia rhodozyma* and is selected from (i) a DNA sequence represented in SEQ ID NOs: 1, 2, 4 or 5; (ii) an isocoding or an allelic variant for the DNA sequence represented in SEQ ID NOs: 1, 2, 4 or 5; and (iii) a derivative of a DNA sequence represented in SEQ ID NOs: 1, 2, 4 or 5 with addition, insertion, deletion and/or substitution of one or more nucleotide(s), and coding for a polypeptide having the said enzyme activity. Such derivatives can be made by recombinant means on the basis of the DNA sequences as disclosed herein by methods known in the state of the art and disclosed e.g. by Sambrook et al. (Molecular Cloning, Cold Spring Harbour Laboratory Press, New York, USA, second edition 1989). Amino acid exchanges in proteins and peptides which do not generally alter the activity are known in the state of the art and are described, for example, by H. Neurath and R. L. Hill in *The Proteins* (Academic Press, New York, 1979, see especially Figure 6, page 14). The most commonly occurring exchanges are: Ala/Ser, Val/Ile, Asp/Glu, Thr/Ser, Ala/Gly, Ala/Thr, Ser/Asn, Ala/Val, Ser/Gly, Tyr/Phe, Ala/Pro, Lys/Arg, Asp/Asn, Leu/Ile, Leu/Val, Ala/Glu, Asp/Gly, as well as these in reverse.

[0008] The present invention also provides an isolated DNA sequence, which is selected from (i) a DNA sequence represented in SEQ ID NO: 3; (ii) an isocoding or an allelic variant for the DNA sequence represented in SEQ ID NO: 3; and (iii) a derivative of a DNA sequence represented in SEQ ID NO: 3 with addition, insertion, deletion and/or substitution of one or more nucleotide(s), and coding for a polypeptide having the mevalonate kinase activity.

[0009] Furthermore the present invention is directed to those DNA sequences as specified above and as disclosed, e.g. in the sequence listing as well as their complementary strands, or those which include these sequences, DNA sequences which hybridize under standard conditions with such sequences or fragments thereof and DNA sequences, which because of the degeneration of the genetic code, do not hybridize under standard conditions with such sequences but which code for polypeptides having exactly the same amino acid sequence.

[0010] "Standard conditions" for hybridization mean in this context the conditions which are generally used by a man skilled in the art to detect specific hybridization signals and which are described, e.g. by Sambrook et al., "Molecular Cloning" second edition, Cold Spring Harbour Laboratory Press 1989, New York, or preferably so called stringent hybridization and non-stringent washing conditions or more preferably so called stringent hybridization and stringent washing conditions a man skilled in the art is familiar with and which are described, e.g. in Sambrook et al. (s.a.). Furthermore DNA sequences which can be made by the polymerase chain reaction by using primers designed on the basis of the DNA sequences disclosed herein by methods known in the art are also an object of the present invention. It is understood that the DNA sequences of the present invention can also be made synthetically as described, e.g. in EP 747 483.

[0011] Further provided by the present invention is a recombinant DNA, preferably a vector and/or plasmid comprising a sequence coding for an enzyme functional in the mevalonate pathway or the reaction pathway from isopentenyl pyrophosphate to farnesyl pyrophosphate. The said recombinant DNA vector and/or plasmid may comprise the regulatory regions such as promoters and terminators as well as open reading frames of above named DNAs.

[0012] The present invention also provides the use of the said recombinant DNA, vector or plasmid, to transform a host organism. The recombinant organism obtained by use of the recombinant DNA is capable of overexpressing DNA sequence encoding an enzyme involved in the mevalonate pathway or the reaction pathway from isopentenyl pyrophosphate to farnesyl pyrophosphate. The host organism transformed with the recombinant DNA may be useful in the improvement of the production process of isoprenoids and carotenoids, in particular astaxanthin. Thus the present invention also provides such a recombinant organism/transformed host.

[0013] The present invention further provides a method for the production of isoprenoids or carotenoids, preferably carotenoids, which comprises cultivating thus obtained recombinant organism.

[0014] The present invention also relates to a method for producing an enzyme involved in the mevalonate pathway or the reaction pathway from isopentenyl pyrophosphate to farnesyl pyrophosphate, which comprises culturing a recombinant organism mentioned above, under a condition conducive to the production of said enzyme and relates also to the enzyme itself.

[0015] The present invention will be understood more easily on the basis of the enclosed figures and the more detailed explanations given below.

Fig. 1 depicts a scheme of deduced biosynthetic pathway from acetyl-CoA to astaxanthin in *P. rhodozyma*.

Fig. 2 shows the expression study by using an artificial *mvk* gene obtained from an artificial nucleotide addition at

amino terminal end of pseudo-*mvk* gene from *P. rhodozyma*. The cells from 50 μ l of broth were subjected to 10 % sodium dodecyl sulfide - polyacrylamide gel electrophoresis (SDS-PAGE). Lane 1, *E. coli* (M15 (pREP4) (pQE30) without IPTG); Lane 2, *E. coli* (M15 (pREP4) (pQE30) with 1mM IPTG); Lane 3, Molecular weight marker (105 kDa, 82.0 kDa, 49.0 kDa, 33.3 kDa and 28.6 kDa, up to down, BIO-RAD); Lane 4, *E. coli* (M15 (pREP4) (pMK1209 #3334) without IPTG); Lane 5, *E. coli* (M15 (pREP4) (pMK1209 #3334) with 1mM IPTG).

[0016] The present invention provides an isolated DNA sequence which code for enzymes which are involved in a biological pathway comprising the mevalonate pathway or the reaction pathway from isopentenyl pyrophosphate to farnesyl pyrophosphate. The said enzymes can be exemplified by those involved in the mevalonate pathway or the reaction pathway from isopentenyl pyrophosphate to farnesyl pyrophosphate in *Phaffia rhodozyma*, such as 3-hydroxy-3-methylglutaryl-CoA synthase, 3-hydroxy-3-methylglutaryl-CoA reductase, mevalonate kinase, mevalonate pyrophosphate decarboxylase and farnesyl pyrophosphate synthase. The present invention is useful for the production of the compounds involved in the biological pathway from the mevalonate pathway to the carotenogenic pathway and various products derived from such compounds. The compounds involved in the mevalonate pathway are acetoacetyl-CoA, 3-hydroxymethyl-3-glutaryl-CoA, mevalonic acid, mevalonate-phosphate, mevalonate-pyrophosphate and isopentenyl-pyrophosphate. Subsequently, isopentenyl-pyrophosphate is converted to geranylgeranyl-pyrophosphate through geranyl-pyrophosphate and farnesyl-pyrophosphate via the "Isoprene Biosynthesis" reactions as indicated in Fig. 1. The compounds involved in the carotenogenic pathway are geranylgeranyl-pyrophosphate, phytoene, lycopene, β -carotene and astaxanthin. Among the compounds involved in the above-mentioned biosynthesis, geranyl-pyrophosphate may be utilized for the production of ubiquinone. Farnesyl-pyrophosphate can be utilized for the production of sterols, such as cholesterol and ergosterol. Geranylgeranyl-pyrophosphate is a useful material for the production of vitamin K, vitamin E, chlorophyll and the like. Thus the present invention will be particularly useful when it is applied to a biological production of isoprenoids. Isoprenoids is the general term which collectively designates a series of compounds having isopentenyl-pyrophosphate as a skeleton unit. Further examples of isoprenoids are vitamin A and vitamin D₃.

[0017] The said DNA of the present invention can mean a cDNA which contains only open reading frame flanked between the short fragments in its 5'- and 3'- untranslated region and a genomic DNA which also contains its regulatory sequences such as its promoter and terminator which are necessary for the expression of the gene of interest.

[0018] In general, the gene consists of several parts which have different functions from each other. In eukaryotes, genes which encode corresponding protein are transcribed to premature messenger RNA (pre-mRNA) differing from the genes for ribosomal RNA (rRNA), small nuclear RNA (snRNA) and transfer RNA (tRNA). Although RNA polymerase II (PolII) plays a central role in this transcription event, PolII can not solely start transcription without *cis* element covering an upstream region containing a promoter and an upstream activation sequence (UAS), and a *trans*-acting protein factor. At first, a transcription initiation complex which consists of several basic protein components recognize the promoter sequence in the 5'-adjacent region of the gene to be expressed. In this event, some additional participants are required in the case of the gene which is expressed under some specific regulation, such as a heat shock response, or adaptation to a nutrition starvation, and so on. In such a case, a UAS is required to exist in the 5'-untranslated upstream region around the promoter sequence, and some positive or negative regulator proteins recognize and bind to the UAS. The strength of the binding of transcription initiation complex to the promoter sequence is affected by such a binding of the *trans*-acting factor around the promoter, and this enables the regulation of the transcription activity.

[0019] After the activation of a transcription initiation complex by the phosphorylation, a transcription initiation complex initiates transcription from the transcription start site. Some parts of the transcription initiation complex are detached as an elongation complex from the promoter region to the 3' direction of the gene (this step is called as a promoter clearance event) and an elongation complex continues the transcription until it reaches to a termination sequence that is located in the 3'-adjacent downstream region of the gene. Pre-mRNA thus generated is modified in nucleus by the addition of cap structure at the cap site which almost corresponds to the transcription start site, and by the addition of polyA stretches at the polyA signal which locates at the 3'-adjacent downstream region. Next, intron structures are removed from coding region and exon parts are combined to yield an open reading frame whose sequence corresponds to the primary amino acid sequence of a corresponding protein. This modification in which a mature mRNA is generated is necessary for a stable gene expression. cDNA in general terms corresponds to the DNA sequence which is reverse-transcribed from this mature mRNA sequence. It can be synthesized by the reverse transcriptase derived from viral species by using a mature mRNA as a template, experimentally.

[0020] To express a gene which was derived from eukaryote, a procedure in which cDNA is cloned into an expression vector in *E. coli* is often used as shown in this invention. This causes from a fact that a specificity of intron structure varies among the organisms and an inability to recognize the intron sequence from other species. In fact, prokaryote has no intron structure in its own genetic background. Even in the yeast, genetic background is different between ascomycetea to which *Saccharomyces cerevisiae* belongs and basidiomycetea to which *P. rhodozyma* belongs. Wery *et al.* showed that the intron structure of actin gene from *P. rhodozyma* cannot be recognized nor spliced by the ascomycetous yeast, *Saccharomyces cerevisiae* (Yeast, 12, 641-651, 1996).

genomic fragment encoding the enzyme of interest into the appropriate vector on which a selectable marker that functions in *P. rhodozyma* is harbored. A drug resistance gene which encodes the enzyme that enables the host survive in the presence of a toxic antibiotic is often used for the selectable marker. G418 resistance gene harbored in pGB-Ph9 (Wery *et al.* (Gene, 184, 89-97, 1997)) is an example of a drug resistance gene. Nutrition complementation marker can be also used in the host which has an appropriate auxotrophy marker. *P. rhodozyma* ATCC24221 strain which requires cytidine for its growth is one example of the auxotroph. By using CTP synthetase as donor DNA for ATCC24221, a host vector system using a nutrition complementation can be established. As a vector, two types of vectors would be used. One of the vectors is an integrated vector which does not have an autonomous replicating sequence. Above pGB-Ph9 is an example of this type of a vector. Because such a vector does not have an autonomous replicating sequence in the vector, above vector cannot replicate by itself and can be present only in an integrated form on the chromosome of the host as a result of a single-crossing recombination using the homologous sequence between a vector and a chromosome. In case of increasing a dose of the integrated gene on the chromosome, amplification of the gene is often employed by using such a drug resistance marker. By increasing the concentration of the corresponding drug in the selection medium, the strain, in which the integrated gene is amplified on the chromosome as a result of recombination only can survive. By using such a selection, a strain which has amplified gene can be chosen. Another type of vector is a replicable vector which has an autonomous replicating sequence. Such a vector can exist in a multicopy state and this makes a dose of the harbored gene also exist in a multicopy state. By using such a strategy, an enzyme of interest which is coded by the amplified gene is expected to be overexpressed.

[0031] Another strategy to overexpress an enzyme of interest is a placement of a gene of interest under a strong promoter. In such a strategy, a copy number of a gene is not necessary to be in a multicopy state. This strategy is also applied to overexpress a gene of interest under the appropriate promoter whose promoter activity is induced in an appropriate growth phase and an appropriate timing of cultivation. Production of astaxanthin accelerates in a late phase of the growth such as the case of production of a secondary metabolite. Thus, the expression of carotenogenic genes may be maximized in a late phase of growth. In such a phase, gene expression of most biosynthesis enzyme decreases. For example, by placing a gene, which is involved in the biosynthesis of a precursor of astaxanthin and whose expression is under the control of a vegetative promoter such as a gene which encodes an enzyme which involves in mevalonate pathway, in the downstream of the promoter of carotenogenic genes, all the genes which are involved in the biosynthesis of astaxanthin become synchronized in their timings and phases of expression.

[0032] Still another strategy to overexpress enzymes of interest is induction of the mutation in its regulatory elements. For this purpose, a kind of reporter gene such as β -galactosidase gene, luciferase gene, a gene coding a green fluorescent protein, and the like is inserted between the promoter and the terminator sequence of the gene of interest so that all the parts including promoter, terminator and the reporter gene are fused and function each other. Transformed *P. rhodozyma* in which the said reporter gene is introduced on the chromosome or on the vector would be mutagenized *in vivo* to induce mutation within the promoter region of the gene of interest. Mutation can be monitored by detecting the change of the activity coded by the reporter gene. If the mutation occurs in a *cis* element of the gene, mutation point would be determined by the rescue of the mutagenized gene and sequencing. The determined mutation would be introduced to the promoter region on the chromosome by the recombination between a native promoter sequence and a mutated sequence. In the same procedure, the mutation occurring in the gene which encodes a *trans*-acting factor can be also obtained. It would also affect the overexpression of the gene of interest.

[0033] A mutation can be also induced by an *in vitro* mutagenesis of a *cis* element in the promoter region. In this approach, a gene cassette, containing a reporter gene which is fused to a promoter region derived from a gene of interest at its 5'-end and a terminator region from a gene of interest at its 3'-end, is mutagenized and then introduced into *P. rhodozyma*. By detecting the difference of the activity of the reporter gene, an effective mutation would be screened. Such a mutation can be introduced in the sequence of the native promoter region on the chromosome by the same method as the case of an *in vivo* mutation approach.

[0034] As a donor DNA, a gene which encodes an enzyme of mevalonate pathway or FPP synthase could be introduced solely or co-introduced by harboring on plasmid vector. A coding sequence which is identical to its native sequence, as well as its allelic variant, a sequence which has one or more amino acid additions, deletions and/or substitutions can be used as far as its corresponding enzyme has the stated enzyme activity. And such a vector can be introduced into *P. rhodozyma* by transformation and a transformant can be selected by spreading the transformed cells on an appropriate selection medium such as YPD agar medium containing geneticin in the case of pGB-Ph9 as a vector or a minimal agar medium omitting cytidine in the case of using auxotroph ATCC24221 as a recipient.

[0035] Such a genetically engineered *P. rhodozyma* would be cultivated in an appropriate medium and evaluated in its productivity of astaxanthin. A hyper producer of astaxanthin thus selected would be confirmed in view of the relationship between its productivity and the level of gene or protein expression which is introduced by such a genetic engineering method.

[0021] Some other researchers reported that intron structures of some kinds of the genes involve regulation of their gene expressions (Dabeva, M. D. *et al.*, Proc. Natl. Acad. Sci. U.S.A., 83, 5854, 1986). It might be important to use a genomic fragment which has its introns in a case of self-cloning of the gene of a interest whose intron structure involves such a regulation of its own gene expression.

5 [0022] To apply a genetic engineering method for a strain improvement study, it is necessary to study its genetic mechanism in the event such as transcription and translation. It is important to determine a genetic sequence such as its UAS, promoter, intron structure and terminator to study the genetic mechanism.

[0023] According to this invention, the genes which code for the enzymes involving the mevalonate pathway were cloned from genomic DNA of *P. rhodozyma*, and their genomic sequence containing HMG-CoA synthase (*hmc*) gene, 10 HMG-CoA reductase (*hmg*) gene, mevalonate kinase (*mvk*) gene, mevalonate pyrophosphate decarboxylase (*mpd*) gene and FPP synthase (*fps*) gene including their 5'- and 3'-adjacent regions as well as their intron structures were determined.

[0024] At first, we cloned a partial gene fragment containing a portion of *hmc* gene, *hmg* gene, *mvk* gene, *mpd* gene and *fps* gene by using degenerate PCR method. The said degenerate PCR is a method to clone a gene of interest 15 which has high homology of amino acid sequence to the known enzyme from other species which has a same or similar function. Degenerate primer, which is used as a primer in degenerate PCR, was designed by a reverse translation of the amino acid sequence to corresponding nucleotides („degenerated”). In such a degenerate primer, a mixed primer which consists any of A, C, G or T, or a primer containing inosine at an ambiguity code is generally used. In this invention, such the mixed primers were used for degenerate primers to clone above genes. PCR condition used is varied 20 depending on primers and genes to clone as described hereinafter.

[0025] An entire gene containing its coding region with its intron as well as its regulation region such as a promoter or terminator can be cloned from a chromosome by screening of genomic library which is constructed in phage vector or plasmid vector in an appropriate host, by using a partial DNA fragment obtained by degenerate PCR as described above as a probe after it was labeled. Generally, *E. coli* as a host strain and *E. coli* vector, a phage vector such as λ 25 phage vector, or a plasmid vector such as pUC vector is often used in the construction of library and a following genetic manipulation such as a sequencing, a restriction digestion, a ligation and the like. In this invention, an *EcoRI* genomic library of *P. rhodozyma* was constructed in the derivatives of λ vector, λ ZAPII and λ DASHII depending on an insert size. An insert size, what length of insert must be cloned, was determined by the Southern blot hybridization for each gene before a construction of a library. In this invention, a DNA which was used for a probe was labeled with digoxigenin (DIG), a steroid hapten instead of conventional ^{32}P label, following the protocol which was prepared by the supplier (Boehringer-Mannheim). A genomic library constructed from the chromosome of *P. rhodozyma* was screened by using 30 a DIG-labeled DNA fragment which had a portion of a gene of interest as a probe. Hybridized plaques were picked up and used for further study. In the case of using λ DASHII (insert size was from 9 kb to 23 kb), prepared λ DNA was digested by the *EcoRI*, followed by the cloning of the *EcoRI* insert into a plasmid vector such as pUC19 or pBluescriptII SK+. When λ ZAPII was used in the construction of the genomic library, *in vivo* excision protocol was conveniently used for the succeeding step of the cloning into the plasmid vector by using a derivative of single stranded M13 phage, Ex assist phage (Stratagene). A plasmid DNA thus obtained was examined for a sequencing. 35

[0026] In this invention, we used the automated fluorescent DNA sequencer, ALFred system (Pharmacia) using an autocycle sequencing protocol in which the Taq DNA polymerase is employed in most cases of sequencing.

40 [0027] After the determination of the genomic sequence, a sequence of a coding region was used for a cloning of cDNA of corresponding gene. The PCR method was also exploited to clone cDNA fragment. The PCR primers whose sequences were identical to the sequence at the 5'- and 3'- end of the open reading frame (ORF) were synthesized with an addition of an appropriate restriction site, and PCR was performed by using those PCR primers. In this invention, a cDNA pool was used as a template in this PCR cloning of cDNA. The said cDNA pool consists of various cDNA species 45 which were synthesized *in vitro* by the viral reverse transcriptase and Taq polymerase (CapFinder Kit manufactured by Clontech was used) by using the mRNA obtained from *P. rhodozyma* as a template. cDNA of interest thus obtained was confirmed in its sequence. Furthermore, cDNA thus obtained was used for a confirmation of its enzyme activity after the cloning of the cDNA fragment into an expression vector which functions in *E. coli* under the strong promoter activity such as the *lac* or T7 expression system.

50 [0028] Succeeding to the confirmation of the enzyme activity, an expressed protein would be purified and used for raising of the antibody against the purified enzyme. Antibody thus prepared would be used for a characterization of the expression of the corresponding enzyme in a strain improvement study, an optimization study of the culture condition, and the like.

[0029] After the rate-limiting step is determined in the biosynthetic pathway which consists of multiple steps of enzymatic reactions, there are three strategies to enhance its enzymatic activity of the rate-limiting reaction by using its 55 genomic sequence.

[0030] One strategy is to use its gene itself as a native form. The simplest approach is to amplify the genomic sequence including its regulation sequence such as a promoter and a terminator. This is realized by the cloning of the

Examples

[0036] The following materials and methods were employed in the Example described below:

5 Strains

[0037] *P. rhodozyma* ATCC96594 (This strain has been redeposited on April 8, 1998 as a Budapest Treaty deposit under accession No. 74438).

10 *E. coli* DH5 α : F⁻, ϕ 80d, *lacZ* Δ M15, Δ (*lacZYA-argF*)U169, *hsd* (*r*_K⁻, *m*_K⁺), *recA1*, *endA1*, *deoR*, *thi-1*, *supE44*, *gyrA96*, *relA1* (Toyobo)

E. coli XL1-Blue MRF': Δ (*mcrA*)183, Δ (*mcrCB-hsdSMR-mrr*)173, *endA1*, *supE44*, *thi-1*, *recA1*, *gyrA96*, *relA1*, *lac*[F' *proAB*, *lac*^PZ Δ M15, Tn10 (*tet*^r)] (Stratagene)

15 *E. coli* SOLR: e14'(*mcrA*), Δ (*mcrCB-hsdSMR-mrr*)171, *sbcC*, *recB*, *recJ*, *umuC* :: Tn5(*kan*^r), *uvrC*, *lac*, *gyrA96*, *relA1*, *thi-1*, *endA1*, λ ^R, [F' *proAB*, *lac*^PZ Δ M15] Su⁻(nonsuppressing) (Stratagene, CA, USA)

20 *E. coli* XL1 MRA (P2): Δ (*mcrA*)183, Δ (*mcrCB-hsdSMR-mrr*)173, *endA1*, *supE44*, *thi-1*, *gyrA96*, *relA1*, *lac* (P2 lysogen) (Stratagene)

E. coli BL21 (DE3) (pLysS): *dcm*⁻, *ompT*_B⁻ *mB*⁻ *lon*⁻ λ (DE3), pLysS (Stratagene)

25 *E. coli* M15 (pREP4) (QIAGEN) (Zamenhof P. J. *et al.*, J. Bacteriol. 110, 171-178, 1972)

E. coli KB822: *pcnB80*, *zad* :: Tn10, Δ (*lacU*169), *hsdR17*, *endA1*, *thi-1*, *supE44*

30 *E. coli* TOP10: F⁻, *mcrA*, Δ (*mrr-hsdRMS-mcrBC*), ϕ 80, Δ *lacZ* M15, Δ *lacX74*, *recA1*, *deoR*, *araD139*, (*ara-leu*)7697, *galU*, *galK*, *rpsL* (Str^r), *endA1*, *nupG* (Invitrogen)

Vectors

[0038]

35 λ ZAPII (Stratagene)

λ DASHII (Stratagene)

pBluescriptII SK+ (Stratagene)

40 pUC57 (MBI Fermentas)

pMOSBlue T-vector (Amersham)

45 pET4c (Stratagene)

pQE30 (QIAGEN)

50 pCR2.1TOPO (Invitrogen)

Media

[0039] *P. rhodozyma* strain is maintained routinely in YPD medium (DIFCO). *E. coli* strain is maintained in LB medium (10 g Bacto-trypton, 5 g yeast extract (DIFCO) and 5 g NaCl per liter). NZY medium (5 g NaCl, 2 g MgSO₄·7H₂O, 5 g yeast extract (DIFCO), 10 g NZ amine type A (Sheffield) per liter) is used for λ phage propagation in a soft agar (0.7 % agar (WAKO)). When an agar medium was prepared, 1.5 % of agar (WAKO) was supplemented.

Methods

[0040] General methods of molecular genetics were practiced according to Molecular cloning: a Laboratory Manual, 2nd Edition (Cold Spring Harbor Laboratory Press, 1989). Restriction enzymes and T4 DNA ligase were purchased from Takara Shuzo (Japan).

[0041] Isolation of a chromosomal DNA from *P. rhodozyma* was performed by using QIAGEN Genomic Kit (QIAGEN) following the protocol supplied by the manufacturer. Mini-prep of plasmid DNA from transformed *E. coli* was performed with the Automatic DNA isolation system (PI-50, Kurabo, Co. Ltd., Japan). Midi-prep of plasmid DNA from an *E. coli* transformant was performed by using QIAGEN column (QIAGEN). Isolation of λ DNA was performed by Wizard lambda preps DNA purification system (Promega) following the protocol of the manufacturer. A DNA fragment was isolated and purified from agarose by using QIAquick or QIAEX II (QIAGEN). Manipulation of λ phage derivatives was done according to the protocol of the manufacturer (Stratagene).

[0042] Isolation of total RNA from *P. rhodozyma* was performed by the phenol method using Isogen (Nippon Gene, Japan). mRNA was purified from total RNA thus obtained by using mRNA separation kit (Clontech). cDNA was synthesized by using CapFinder cDNA construction kit (Clontech).

[0043] *In vitro* packaging was performed by using Gigapack III gold packaging extract (Stratagene).

[0044] Polymerase chain reaction (PCR) is performed with the thermal cycler from Perkin Elmer model 2400. Each PCR condition is described in examples. PCR primers were purchased from a commercial supplier or synthesized with a DNA synthesizer (model 392, Applied Biosystems). Fluorescent DNA primers for DNA sequencing were purchased from Pharmacia. DNA sequencing was performed with the automated fluorescent DNA sequencer (ALFred, Pharmacia).

[0045] Competent cells of DH5 α were purchased from Toyobo (Japan). Competent cells of M15 (pREP4) were prepared by CaCl₂ method as described by Sambrook *et al.* (Molecular cloning: a Laboratory Manual, 2nd Edition, Cold Spring Harbor Laboratory Press, 1989).

Example 1 Isolation of mRNA from *P. rhodozyma* and construction of cDNA library

[0046] To construct cDNA library of *P. rhodozyma*, total RNA was isolated by phenol extraction method right after the cell disruption and the mRNA from *P. rhodozyma* ATCC96594 strain was purified by using mRNA separation kit (Clontech).

[0047] At first, Cells of ATCC96594 strain from 10 ml of two-day-culture in YPD medium were harvested by centrifugation (1500 x g for 10 min.) and washed once with extraction buffer (10 mM Na-citrate / HCl (pH 6.2) containing 0.7 M KCl). After suspending in 2.5 ml of extraction buffer, the cells were disrupted by French press homogenizer (Ohtake Works Corp., Japan) at 1500 kgf/cm² and immediately mixed with two times of volume of isogen (Nippon gene) according to the method specified by the manufacturer. In this step, 400 μ g of total RNA was recovered.

[0048] Then this total RNA was purified by using mRNA separation kit (Clontech) according to the method specified by the manufacturer. Finally, 16 μ g of mRNA from *P. rhodozyma* ATCC96594 strain was obtained.

[0049] To construct cDNA library, CapFinder PCR cDNA construction kit (Clontech) was used according to the method specified by the manufacturer. One μ g of purified mRNA was applied for a first strand synthesis followed by PCR amplification. After this amplification by PCR, 1 mg of cDNA pool was obtained.

Example 2 Cloning of the partial *hmc* (3-hydroxy-3-methylglutaryl-CoA synthase) gene from *P. rhodozyma*

[0050] To clone a partial *hmc* gene from *P. rhodozyma*, a degenerate PCR method was exploited. Two mixed primers whose nucleotide sequences were designed and synthesized as shown in TABLE 1 based on the common sequence of known HMG-CoA synthase genes from other species.

TABLE 1

Sequence of primers used in the cloning of *hmc* gene

Hmgs1 ; GGNAARTAYACNATHGAGNYTNGGNC (sense primer) (SEQ ID NO: 11)

Hmgs3 ; TANARNWSWNGTRTACATRTTNC (antisense primer) (SEQ ID NO: 12)

(N=A, C, G or T; R=A or G, Y=C or T, H=A, T or C, S=C or G, W=A or T)

[0051] After the PCR reaction of 25 cycles of 95 °C for 30 seconds, 50 °C for 30 seconds and 72°C for 15 seconds by using ExTaq (Takara Shuzo) as a DNA polymerase and cDNA pool obtained in example 1 as a template, reaction

mixture was applied to agarose gel electrophoresis. A PCR band that has a desired length was recovered and purified by QIAquick (QIAGEN) according to the method by the manufacturer and then ligated to pMOSBlue T-vector (Amersham). After the transformation of competent *E. coli* DH5 α , 6 white colonies were selected and plasmids were isolated with Automatic DNA isolation system. As a result of sequencing, it was found that 1 clone had a sequence whose deduced amino acid sequence was similar to known *hmc* genes. This isolated cDNA clone was designated as pHMC211 and used for further study.

Example 3 Isolation of genomic DNA from *P. rhodozyma*

[0052] To isolate a genomic DNA from *P. rhodozyma*, QIAGEN genomic kit was used according to the method specified by the manufacturer.

[0053] At first, cells of *P. rhodozyma* ATCC96594 strain from 100 ml of overnight culture in YPD medium were harvested by centrifugation (1500 x g for 10 min.) and washed once with TE buffer (10 mM Tris / HCl (pH 8.0) containing 1 mM EDTA). After suspending in 8 ml of Y1 buffer of the QIAGEN genomic kit, lyticase (SIGMA) was added at the concentration of 2 mg/ml to disrupt cells by enzymatic degradation and the reaction mixture was incubated for 90 minutes at 30 °C and then proceeded to the next extraction step. Finally, 20 μ g of genomic DNA was obtained.

Example 4 Southern blot hybridization by using pHMC211 as a probe

[0054] Southern blot hybridization was performed to clone a genomic fragment which contains *hmc* gene from *P. rhodozyma*. Two μ g of genomic DNA was digested by *EcoRI* and subjected to agarose gel electrophoresis followed by acidic and alkaline treatment. The denatured DNA was transferred to nylon membrane (Hybond N+, Amersham) by using transblot (Joto Rika) for an hour. The DNA which was transferred to nylon membrane was fixed by a heat treatment (80 °C, 90 minutes). A probe was prepared by labeling a template DNA (*EcoRI*- and *SaII*- digested pHMC211) with DIG multipriming method (Boehringer Mannheim). Hybridization was performed with the method specified by the manufacturer. As a result, hybridized band was visualized in the range from 3.5 to 4.0 kilobases (kb).

Example 5 Cloning of a genomic fragment containing *hmc* gene

[0055] Four μ g of the genomic DNA was digested by *EcoRI* and subjected to agarose gel electrophoresis. Then, DNAs whose length is within the range from 3.0 to 5.0 kb was recovered by QIAEX II gel extraction kit (QIAGEN) according to the method specified by the manufacturer. The purified DNA was ligated to 1 μ g of *EcoRI*-digested and CIAP (calf intestine alkaline phosphatase) -treated λ ZAPII (Stratagene) at 16 °C overnight, and packaged by Gigapack III gold packaging extract (Stratagene). The packaged extract was infected to *E. coli* XL1Blue MRF' strain and over-laid with NZY medium poured onto LB agar medium. About 6000 plaques were screened by using *EcoRI*- and *SaII*-digested pHMC211 as a probe. Two plaques were hybridized to the labeled probe and subjected to *in vivo* excision protocol according to the method specified by the manufacturer (Stratagene). It was found that isolated plasmids had the same fragments in the opposite direction each other as results of restriction analysis and sequencing. As a result of sequencing, the obtained *EcoRI* fragment contained same nucleotide sequence as that of pHMC211 clone. One of these plasmids was designated as pHMC526 and used for further study. A complete nucleotide sequence was obtained by sequencing of deletion derivatives of pHMC526, and sequencing with a primer-walking procedure. The insert fragment of pHMC526 consists of 3431 nucleotides that contained 10 complete and an incomplete exons and 10 introns with about 1 kb of 3'-terminal untranslated region.

Example 6 Cloning of upstream region of *hmc* gene

[0056] Cloning of 5'- adjacent region of *hmc* gene was performed by using Genome Walker Kit (Clontech), because pHMC 526 does not contain its 5' end of *hmc* gene. At first, the PCR primers whose sequences were shown in TABLE 2 were synthesized.

TABLE 2

Sequence of primers used in the cloning of 5'- adjacent region of *hmc* gene

Hmc21 ; GAAGAACCCCATCAAAGCCTCGA (primary primer) (SEQ ID NO: 13)

Hmc22 ; AAAAGCCTCGAGATCCTTGTGAGCG (nested primer) (SEQ ID NO: 14)

[0057] Protocols for library construction and PCR condition were the same as those specified by the manufacturer by using the genomic DNA preparation obtained in Example 3 as a PCR template. The PCR fragments that had *EcoRV* site at the 5' end (0.45 kb), and that had *PvuII* site at the 5' end (2.7 kb) were recovered and cloned into pMOSBlue T-vector by using *E. coli* DH5 α as a host strain. As a result of sequencing of each 5 of independent clones from both constructs, it was confirmed that the 5' adjacent region of *hmc* gene was cloned and small part (0.1 kb) of *EcoRI* fragment within its 3' end was found. The clone obtained by the *PvuII* construct in the above experiment was designated as pHMCPv708 and used for further study.

[0058] Next, Southern blot analysis was performed by the method as shown in above Example 4, and 5'- adjacent region of the *hmc* gene existed in 3 kb of *EcoRI* fragment was determined. After a construction from 2.5 to 3.5 kb *EcoRI* library in λ ZAPII, 600 plaques were screened and 6 positive clones were selected. As a result of sequencing of these 6 clones, it was clarified that 4 clones within 6 positive plaques had the same sequence as that of the pHMCPv708, and one of those was named as pHMC723 and used for further analysis.

[0059] The PCR primers whose sequences were shown in TABLE 3 were synthesized to clone small (0.1 kb) *EcoRI* fragment locating between 3.5 kb and 3.0 kb *EcoRI* fragments on the chromosome of *P. rhodozyma*.

TABLE 3

Sequence of primers used in the cloning small *EcoRI* portion of *hmc* gene.

Hmc30 ; AGAAGCCAGAAGAGAAAA (sense primer) (SEQ ID NO: 15)

Hmc31 ; TCGTCGAGGAAAGTAGAT (antisense primer) (SEQ ID NO: 16)

[0060] The PCR condition was the same as shown in Example 2. Amplified fragment (0.1 kb in its length) was cloned into pMOSBlue T-vector and transformed *E. coli* DH5 α . Plasmids were prepared from 5 independent white colonies and subjected to the sequencing.

[0061] Thus, it was determined that the nucleotide sequence (4.8 kb) contained *hmc* gene (SEQ ID NO: 1). Coding region was in 2432 base pairs that consisted of 11 exons and 10 introns. Introns were scattered all through the coding region without 5' or 3' bias. It was found that open reading frame consists of 467 amino acids (SEQ ID NO: 6) whose sequence is strikingly similar to the known amino acid sequence of HMG-CoA synthase gene from other species (49.6 % identity to HMG-CoA synthase from *Schizosaccharomyces pombe*).

Example 7 Expression of *hmc* gene in *E. coli* and confirmation of its enzymatic activity

[0062] The PCR primers whose sequences were shown in TABLE 4 were synthesized to clone a cDNA fragment of *hmc* gene.

TABLE 4

Sequence of primers used in the cloning of cDNA of *hmc* gene

Hmc25 ; GGTACCATATGTATCCTTCTACTACCGAAC (sense primer) (SEQ ID NO: 17)

Hmc26 ; GCATGCGGATCCTCAAGCAGAAGGGACCTG (antisense primer) (SEQ ID NO: 18)

[0063] PCR condition was as follows; 25 cycles of 95 °C for 30 seconds, 55 °C for 30 seconds and 72 °C for 3 minutes. As a template, 0.1 μ g of cDNA pool obtained in Example 2 was used, and Pfu polymerase as a DNA polymerase. Amplified 1.5 kb fragment was recovered and cloned in pT7Blue-3 vector (Novagen) by using perfectly blunt cloning kit (Novagen) according to the protocol specified by the manufacturer. Six independent clones from white colonies of *E. coli* DH5 α transformants were selected and plasmids were prepared from those transformants. As a result of restriction analysis, 2 clones were selected for a further selection by sequencing. One clone has an amino acid substitution at position 280 (from glycine to alanine) and another has at position 53 (from alanine to threonine). Alignment of an amino acid sequences derived from known *hmc* genes showed that alanine residue as well as glycine residue at position 280 was observed well in all the sequences from other species and this fact suggested that amino acid substitution at position 280 would not affect its enzymatic activity. This clone (mutant at position 280) was selected as pHMC731 for a succeeding expression experiment.

[0064] Next, 1.5 kb fragment obtained by *NdeI*- and *BamHI*- digestion of pHMC731 was ligated to pET11c (Stratagene) digested by the same pairs of restriction enzymes, and introduced to *E. coli* DH5 α . As a result of restriction anal-

ysis, plasmid that had a correct structure (pHMC818) was recovered. Then, competent *E. coli* BL21 (DE3) (pLysS) cells (Stratagene) were transformed, and one clone that had a correct structure was selected for further study.

[0065] For an expression study, strain BL21 (DE3) (pLysS) (pHMC818) and vector control strain BL21 (DE3) (pET11c) were cultivated in 100 ml of LB medium at 37 °C until OD at 600 nm reached to 0.8 (about 3 hours) in the presence of 100 µg/ml of ampicillin. Then, the broth was divided in two portions of the same volume, and then 1 mM of isopropyl β-D-thiogalactopyranoside (IPTG) was added to one portion. Cultivation was continued for further 4 hours at 37 °C. Twenty five µl of broth was removed from induced- and uninduced- culture of *hmc* clone and vector control cultures and subjected to sodium dodecyl sulfate - polyacrylamide gel electrophoresis (SDS-PAGE) analysis. It was confirmed that protein whose size was similar to deduced molecular weight from nucleotide sequence (50.8 kDa) was expressed only in the case of clone that harbored pHMC818 with the induction. Cells from 50 ml broth were harvested by the centrifugation (1500 x g, 10 minutes), washed once and suspended in 2 ml of hmc buffer (200 mM Tris-HCl (pH 8.2)). Cells were disrupted by French press homogenizer (Ohtake Works) at 1500 kgf/cm² to yield a crude lysate. After the centrifugation of the crude lysate, a supernatant fraction was recovered and used as a crude extract for an enzymatic analysis. In the only case of induced lysate of pHMC818 clone, a white pellet was spun down and was recovered. Enzyme assay for 3-hydroxy-3-methylglutaryl-CoA (HMG-CoA) synthase was performed by the photometric assay according to the method by Stewart *et al.* (J. Biol. Chem. 241(5), 1212-1221, 1966). In all the crude extract, the activity of 3-hydroxy-3-methylglutaryl-CoA synthase was not detected. As a result of SDS-PAGE analysis of the crude extract, an expressed protein band that had found in expressed broth was disappeared. Subsequently the white pellet that was recovered from the crude lysate of induced pHMC818 clone was solubilized with 8 M guanidine-HCl, and then subjected to SDS-PAGE analysis. The expressed protein was recovered in the white pellet, and this suggested that expressed protein would form an inclusion body.

[0066] Next, an expression experiment in more mild condition was conducted. Cells were grown in LB medium at 28 °C and the induction was performed by the addition of 0.1 mM of IPTG. Subsequently, incubation was continued further for 3.5 hours at 28 °C and then the cells were harvested. Preparation of the crude extract was the same as the previous protocol. Results are summarized in TABLE 5. It was shown that HMG-CoA synthase activity was only observed in the induced culture of the recombinant strain harboring *hmc* gene, and this suggested that the cloned *hmc* gene encodes HMG-CoA synthase.

TABLE 5

Enzymatic characterization of <i>hmc</i> cDNA clone		
plasmid	IPTG	µ mol of HMG-CoA / minute / mg-protein
pHMC818	-	0
	+	0.146
pET11c	-	0
	+	0

Example 8 Cloning of *hmg* (3-hydroxymethyl-3-glutaryl-CoA reductase) gene

[0067] Cloning protocol of *hmg* gene was almost the same as the *hmc* gene shown in Example 2 to 7. At first, the PCR primers whose sequences were shown in TABLE 6 based on the common sequences of HMG-CoA reductase genes from other species were synthesized.

TABLE 6

Sequence of primers used in the cloning of *hmg* gene

Red1 ; GCNTGYTGYGARAAYGTNATHGGNTAYATGCC (sense primer) (SEQ ID NO: 19)

Red2 ; ATCCARTTDATNGCNGCNGGYTTYTTRTCNGT (antisense primer) (SEQ ID NO: 20)

(N=A, C, G or T; R=A or G, Y=C or T, H=A, T or C, D=A, G or T)

[0068] After the PCR reaction of 25 cycles of 95 °C for 30 seconds, 54 °C for 30 seconds and 72°C for 30 seconds by using ExTaq (Takara Shuzo) as a DNA polymerase, reaction mixture was applied to agarose gel electrophoresis.

PCR band that has a desired length was recovered and purified by QIAquick (QIAGEN) according to the method by the manufacturer and then ligated to pUC57 vector (MBI Fermentas). After the transformation of competent *E. coli* DH5 α , 7 white colonies were selected and the plasmids were isolated from those transformants. As a result of sequencing, it was found that all the clones had a sequence whose deduced amino acid sequence was similar to known HMG-CoA reductase genes. One of the isolated cDNA clones was named as pRED1219 and used for further study.

[0069] Next, a genomic fragment containing 5'- and 3'- adjacent region of *hmg* gene was cloned with the Genome Walker kit (Clontech). The 2.5 kb fragment of 5' adjacent region (pREDPVu1226) and 4.0 kb fragment of 3' adjacent region of *hmg* gene (pREDEVd1226) were cloned. Based on the sequence of the insert of pREDPVu1226, PCR primers whose sequence were shown in TABLE 7 were synthesized.

TABLE 7

Sequence of primers used in the cloning of cDNA of *hmg* gene

Red8 ; GGCCATTCCACACTTGATGCTCTGC (antisense primer) (SEQ ID NO: 21)

Red9 ; GGCCGATATCTTTATGGTCCT (sense primer) (SEQ ID NO: 22)

[0070] Subsequently a cDNA fragment containing a long portion of *hmg* cDNA sequence was cloned by a PCR method by using Red 8 and Red 9 as PCR primers and the cDNA pool prepared in Example 2 and thus cloned plasmid was named as pRED107. PCR condition was as follows; 25 cycles of 30 seconds at 94 °C, 30 seconds at 55 °C and 1 minute at 72 °C.

[0071] Southern blot hybridization study was performed to clone genomic sequence which contains the entire *hmg* gene from *P. rhodozyma*. Probe was prepared by labeling a template DNA, pRED107 with DIG multipriming method. Hybridization was performed with the method specified by the manufacturer. As a result, labeled probe hybridized to two bands that had 12 kb and 4 kb in their lengths. As a result of sequencing of pREDPVu1226, *EcoRI* site wasn't found in the cloned *hmg* region. This suggested that another species of *hmg* gene (that has 4 kb of hybridized *EcoRI* fragment) existed on the genome of *P. rhodozyma* as found in other organisms.

[0072] Next, a genomic library consisting of 9 to 23 kb of *EcoRI* fragment in the λ DASHII vector was constructed. The packaged extract was infected to *E. coli* XL1 Blue, MRA(P2) strain (Stratagene) and over-laid with NZY medium poured onto LB agar medium. About 5000 plaques were screened by using 0.6 kb fragment of *StuI*- digested pRED107 as a probe. 4 plaques were hybridized to the labeled probe. Then a phage lysate was prepared and DNA was purified with Wizard lambda purification system according to the method specified by the manufacturer (Promega) and was digested with *EcoRI* to isolate 10 kb of *EcoRI* fragment and to clone in *EcoRI*-digested and CIAP-treated pBluescriptII KS-(Stratagene). Eleven white colonies were selected and subjected to a colony PCR by using Red9 and -40 universal primer (Pharmacia). Template DNA for a colony PCR was prepared by heating cell suspension in which picked-up colony was suspended in 10 μ l of sterilized water for 5 minutes at 99 °C prior to a PCR reaction (PCR condition; 25 cycles of 30 seconds at 94 °C, 30 seconds at 55 °C and 3 minutes at 72 °C). One colony gave 4 kb of a positive PCR band, and it suggested that this clone had an entire region containing *hmg* gene. A plasmid from this positive clone was prepared and named as pRED611. Subsequently deletion derivatives of pRED611 were made up for sequencing. By combining the sequence obtained from deletion mutants with the sequence obtained by a primer-walking procedure, the nucleotide sequence of 7285 base pairs which contains *hmg* gene from *P. rhodozyma* was determined (SEQ ID NO: 2). The *hmg* gene from *P. rhodozyma* consists of 10 exons and 9 introns. The deduced amino acid sequence of 1092 amino acids in its length (SEQ ID NO: 7) showed an extensive homology to known HMG-CoA reductase (53.0 % identity to HMG-CoA reductase from *Ustilago maydis*).

Example 9 Expression of carboxyl-terminal domain of *hmg* gene in *E. coli*

[0073] Some species of prokaryotes have soluble HMG-CoA reductases or related proteins (Lam *et al.*, J. Biol. Chem. 267, 5829-5834, 1992). However, in eukaryotes, HMG-CoA reductase is tethered to the endoplasmic reticulum via an amino-terminal membrane domain (Skalnik *et al.*, J. Biol. Chem. 263, 6836-6841, 1988). In fungi (i.e. *Saccharomyces cerevisiae* and the smut fungus, *Ustilago maydis*) and in animals, the membrane domain is large and complex, containing seven or eight transmembrane segments (Croxen *et al.* Microbiol. 140, 2363-2370, 1994). In contrast, the membrane domains of plant HMG-CoA reductase proteins have only one or two transmembrane segments (Nelson *et al.* Plant Mol. Biol. 25, 401-412, 1994). Despite the difference in the structure and sequence of the transmembrane domain, the amino acid sequences of the catalytic domain are conserved across eukaryotes, archaeobacteria and eubacteria.

[0074] Croxen *et al.* showed that C-terminal domain of HMG-CoA reductase derived from the maize fungal pathogen,

Ustilago maydis was expressed in active form in *E. coli* (Microbiology, 140, 2363-2370, 1994). The inventors of the present invention tried to express a C-terminal domain of HMG-CoA reductase from *P. rhodozyma* in *E. coli* to confirm its enzymatic activity.

[0075] At first, the PCR primers whose sequences were shown in TABLE 8 were synthesized to clone a partial cDNA fragment of *hmg* gene. The sense primer sequence corresponds to the sequence which starts from 597th amino acid (glutamate) residue, and a length of protein and cDNA which was expected to obtain was 496 amino acids and 1.5 kb, respectively.

TABLE 8

Sequence of primers used in the cloning of a partial cDNA of *hmg* gene

Red54 ; GGTACCGAAGAAATTATGAAGAGTGG (sense primer) (SEQ ID NO: 23)

Red55 ; CTGCAGTCAGGCATCCACGTTACAC (antisense primer) (SEQ ID NO: 24)

[0076] The PCR condition was as follows; 25 cycles of 95 °C for 30 seconds, 55 °C for 30 seconds and 72 °C for 3 minutes. As a template, 0.1 µg of cDNA pool obtained in Example 2 and as a DNA polymerase, ExTaq polymerase were used. Amplified 1.5 kb fragment was recovered and cloned in pMOSBlue T-vector (Novagen). Twelve independent clones from white colonies of *E. coli* DH5α transformants were selected and plasmids were prepared from those transformants. As a result of restriction analysis, all the clones were selected for a further selection by sequencing. One clone did not have an amino acid substitution all through the coding sequence and was named as pRED908.

[0077] Next, 1.5 kb fragment obtained by *Kpn*I- and *Pst*I- digestion of pRED908 was ligated to pQE30 (QIAGEN) digested by the same pairs of restriction enzymes, and transformed to *E. coli* KB822. As a result of restriction analysis, plasmid that had a correct structure (pRED1002) was recovered. Then, competent *E. coli* M15 (pREP4) cells (QIAGEN) were transformed and one clone that had a correct structure was selected for further study.

[0078] For an expression study, strain M15 (pREP4) (pRED1002) and vector control strain M15 (pREP4) (pQE30) were cultivated in 100 ml of LB medium at 30 °C until OD at 600 nm reached to 0.8 (about 5 hours) in the presence of 25 µg/ml of kanamycin and 100 µg/ml of ampicillin. Then, the broth was divided into two portions of the same volume, and then 1 mM of IPTG was added to one portion. Cultivation continued for further 3.5 hours at 30 °C. Twenty five µl of the broth was removed from induced- and uninduced- culture of *hmg* clone and vector control cultures and subjected to SDS-PAGE analysis. It was confirmed that protein whose size was similar to deduced molecular weight from nucleotide sequence (52.4 kDa) was expressed only in the case of clone that harbored pRED1002 with the induction. Cells from 50 ml broth were harvested by the centrifugation (1500 x g, 10 minutes), washed once and suspended in 2 ml of *hmg* buffer (100 mM potassium phosphate buffer (pH 7.0) containing 1 mM of EDTA and 10 mM of dithiothreitol). Cells were disrupted by French press (Ohtake Works) at 1500 kgf/cm² to yield a crude lysate. After the centrifugation of the crude lysate, a supernatant fraction was recovered and used as a crude extract for enzymatic analysis. In the only case of induced lysate of pRED1002 clone, a white pellet was spun down and was recovered. Enzyme assay for 3-hydroxy-3-methylglutaryl-CoA (HMG-CoA) reductase was performed by the photometric assay according to the method by Servouse *et al.* (Biochem. J. 240, 541-547, 1986). In all the crude extract, the activity of 3-hydroxy-3-methylglutaryl-CoA synthase was not detected. As a result of SDS-PAGE analysis for the crude extract, expressed protein band that had found in expressed broth was disappeared. Next, the white pellet that was recovered from the crude lysate of induced pRED1002 clone was solubilized with an equal volume of 20 % SDS, and then subjected to SDS-PAGE analysis. An expressed protein was recovered in the white pellet, and this suggested that the expressed protein would form an inclusion body.

[0079] Next, the expression experiment was performed in more mild condition. Cells were grown in LB medium at 28 °C and the induction was performed by the addition of 0.1 mM of IPTG. Then, incubation was continued further for 3.5 hours at 28 °C and then the cells were harvested. Preparation of the crude extract was the same as the previous protocol. Results are summarized in TABLE 9. It was shown that 30 times higher induction was observed, and this suggested that the cloned *hmg* gene codes HMG-CoA reductase.

TABLE 9

Enzymatic characterization of <i>hmg</i> cDNA clone		
plasmid	IPTG μ mol of NADPH / minute / mg-protein	
pRED1002	-	0.002
	+	0.059
pQE30	-	0
	+	0

Example 10 Cloning of mevalonate kinase (*mvk*) gene

[0080] A cloning protocol of *mvk* gene was almost the same as the *hmg* gene shown in Example 2 to 7. At first, the PCR primers whose sequence were shown in TABLE 10, based on the common sequences of mevalonate kinase genes from other species were synthesized.

TABLE 10

Sequence of primers used in the cloning of *mvk* gene

Mk1 ; GCNCCNGGNAARGTNATHYNTTYGGNGA (sense primer) (SEQ ID NO: 25)

Mk2 ; CCCANGTNSWNACNGCRTRTTCNACNCC (antisense primer) (SEQ ID NO: 26)

(N=A, C, G or T; R=A or G, Y=C or T, H=A, T or C, S=C or G, W=A or T)

[0081] After the PCR reaction of 25 cycles of 95 °C for 30 seconds, 46 °C for 30 seconds and 72°C for 15 seconds by using ExTaq as a DNA polymerase, the reaction mixture was applied to agarose gel electrophoresis. A 0.6 kb of PCR band whose length was expected to contain a partial *mvk* gene was recovered and purified by QIAquick according to the method indicated by the manufacturer and then ligated to pMOSBlue T-vector. After a transformation of competent *E. coli* DH5 α cells, 4 white colonies were selected and plasmids were isolated. As a result of sequencing, it was found that one of the clones had a sequence whose deduced amino acid sequence was similar to known mevalonate kinase genes. This cDNA clone was named as pMK128 and used for further study.

[0082] Next, a partial genomic clone which contained *mvk* gene was cloned by PCR. The PCR primers whose sequence were shown in TABLE 11, based on the internal sequence of pMK128 were synthesized.

TABLE 11

Sequence of primers used in the cloning of genomic DNA containing *mvk* gene

Mk5 ; ACATGCTGTAGTCCATG (sense primer) (SEQ ID NO: 27)

Mk6 ; ACTCGGATTCCATGGA (antisense primer) (SEQ ID NOP: 28)

[0083] PCR condition was 25 cycles of 30 seconds at 94 °C, 30 seconds at 55 °C and 1 minute at 72 °C. The amplified 1.4 kb fragment was cloned into pMOSBlue T-vector. As a result of sequencing, it was confirmed a genomic fragment containing *mvk* gene which had typical intron structures could be obtained and this genomic clone was named as pMK224.

[0084] Southern blot hybridization study was performed to clone a genomic fragment which contained an entire *mvk* gene from *P. rhodozyma*. Probe was prepared by labeling a template DNA, pMK224 digested by *Nco*I with DIG multipriming method. Hybridization was performed with the method specified by the manufacturer. As a result, the labeled probe hybridized to a band that had 6.5 kb in its lengths. Next, a genomic library consisting of 5 to 7 kb of *Eco*RI fragment was constructed in the λ ZAPII vector. The packaged extract was infected to *E. coli* XL1Blue, MRF strain (Strata-

gene) and over-laid with NZY medium poured onto LB agar medium. About 5000 plaques were screened by using 0.8 kb fragment of *Nco*I- digested pMK224 as a probe. Seven plaques were hybridized to the labeled probe. Then a phage lysate was prepared according to the method specified by the manufacturer (Stratagene) and *in vivo* excision was performed by using *E. coli* XL1Blue MRF⁺ and SOLR strains. Fourteen white colonies were selected and plasmids were isolated from those selected transformants. Then, isolated plasmids were digested by *Nco*I and subjected to Southern blot hybridization with the same probe as the plaque hybridization. The insert fragments of all the plasmids were hybridized to the probe and this suggested that a genomic fragment containing *mvk* gene could be cloned. A plasmid from one of the positive clones was prepared and was named as pMK701. About 3 kb of sequence was determined by the primer walking procedure and it was revealed that 5' end of the *mvk* gene wasn't included into pMK701.

[0085] Next, a PCR primer which had a sequence ;

TTGTTGTCGTAGCAGTGAGAG (SEQ ID NO: 29) was synthesized to clone the 5'-adjacent genomic region of *mvk* gene with the Genome Walker Kit according to the method specified by manufacturer (Clontech). A specific 1.4 kb PCR band was amplified and cloned into pMOSBlue T-vector. All of the transformants of DH5 α selected had expected length of the insert. Subsequent sequencing revealed that 5'-adjacent region of *mvk* gene could be cloned. One of the clone was designated as pMKEVR715 and used for further study. As a result of Southern blot hybridization using genomic DNA prepared in example 3, the labeled pMKEVR715 hybridized to 2.7 kb *Eco*RI band. Then a genomic library in which *Eco*RI fragments from 1.4 to 3.0 kb in lengths were cloned into λ ZAPII was constructed and screened with 1.0 kb of *Eco*RI fragment from pMKEVR715. Fourteen positive plaques were selected from 5000 plaques and plasmids were prepared from those plaques with *in vivo* excision procedure.

[0086] The PCR primers whose sequences were shown in TABLE 12, taken from the internal sequence of pMKEVR715 were synthesized to select a positive clone with a colony PCR.

TABLE 12

PCR primers used for colony PCR to clone 5'-adjacent region of *mvk* gene

Mk17 ; GGAAGAGGAAGAGAAAAG (sense primer) (SEQ ID NO: 30)

Mk18 ; TTGCCGAACCTCAATGTAG (antisense primer) (SEQ ID NO: 31)

[0087] PCR condition was as follows; 25 cycles of 30 seconds at 94 °C, 30 seconds at 50 °C and 15 seconds at 72 °C. From all the candidates except one clone, the positive 0.5 kb band was yielded. One of the clones was selected and named as pMK723 to determine the sequence of the upstream region of *mvk* gene. After sequencing of the 3'-region of pMK723 and combining with the sequence of pMK701, the genomic sequence of 4.8 kb fragment containing *mvk* gene was determined. The *mvk* gene consists of 4 introns and 5 exons (SEQ ID NO: 3). The deduced amino acid sequence except 4 amino acids in the amino terminal end (SEQ ID NO: 8) showed an extensive homology to known mevalonate kinase (44.3 % identity to mevalonate kinase from *Rattus norvegicus*).

Example 11 Expression of *mvk* gene by the introduction of 1 base at amino terminal region

[0088] Although the amino acid sequence showed a significant homology to known mevalonate kinase, an appropriate start codon for *mvk* gene could not be found. This result suggested the cloned gene might be a pseudogene for mevalonate kinase. To confirm this assumption, PCR primers whose sequences are shown in TABLE 13 were synthesized to introduce an artificial nucleotide which resulted in the generation of appropriate start codon at the amino terminal end.

TABLE 13

PCR primers used for the introduction of a nucleotide into *mvk* gene

Mk33 ; GGATCCATGAGAGCCCAAAAAGAAGA (sense primer) (SEQ ID NO: 32)

Mk34 ; GTCGACTCAAGCAAAAGACCAACGAC (antisense primer) (SEQ ID NO: 33)

[0089] The artificial amino terminal sequence thus introduced were as follows; NH₂-Met-Arg-Ala-Gln. After the PCR reaction of 25 cycles of 95 °C for 30 seconds, 55 °C for 30 seconds and 72 °C for 30 seconds by using ExTaq polymerase as a DNA polymerase. The reaction mixture was subjected to agarose gel electrophoresis. An expected 1.4 kb of PCR band was amplified and cloned into pCR2.1 TOPO vector. After a transformation of competent *E. coli* TOP10 cells,

6 white colonies were selected and plasmids were isolated. As a result of sequencing, it was found that one clone had only one change of amino acid residue (Asp to Gly change at 81st amino acid residue in SEQ ID NO: 8). This plasmid was named as pMK1130 #3334 and used for further study. Then, the insert fragment of pMK1130 #3334 was cloned into pQE30. This plasmid was named as pMK1209 #3334. After the transformation of expression host, M15 (pREP4), expression study was conducted. M15 (pREP4) (pMK1209 #3334) strain and vector control strain (M15 (pREP4) (pQE30)) were inoculated into 3 ml of LB medium containing 100 µg/ml of ampicillin. After the cultivation at 37 °C for 3.75 hours, cultured broth were divided into two portions. 1 mM IPTG were added to one portion and an incubation was continued for 3 hours. Cells were harvested from 50 µl of broth by the centrifugation and were subjected to SDS-PAGE analysis. Protein which had an expected molecular weight of 48.5 kDa was induced by the addition of IPTG in the culture of M15 (pREP4) (pMK1209 #3334) though no induced protein band was observed in the vector control culture (Fig. 2). This result suggested that activated form of the mevalonate kinase protein could be expressed by the artificial addition of one nucleotide at amino terminal end.

Example 12 Cloning of the mevalonate pyrophosphate decarboxylase (*mpd*) gene

[0090] A cloning protocol of *mpd* gene was almost the same as the *hmc* gene shown in Example 2 to 7. At first, the PCR primers whose sequence were shown in TABLE 14 based on the common sequences of mevalonate pyrophosphate decarboxylase genes from other species were synthesized.

TABLE 14

Sequence of primers used in the cloning of *mpd* gene

Mpd1 ; HTNAARTAYTTGGNAARMGNGA (sense primer) (SEQ ID NO: 34)
Mpd2 ; GCRTTNGGNCNGCRTCRAANGTRTANGC (antisense primer) (SEQ ID NO: 35)
(N=A, C, G or T; R=A or G, Y=C or T, H=A, T or C, M=A or C)

[0091] After the PCR reaction of 25 cycles of 95 °C for 30 seconds, 50 °C for 30 seconds and 72°C for 15 seconds by using ExTaq as a DNA polymerase, reaction mixture was subjected to agarose gel electrophoresis. A 0.9 kb of PCR band whose length was expected to contain a partial *mpd* gene was recovered and purified by QIAquick according to the method prepared by the manufacturer and then ligated to pMOSBlue T-vector. After a transformation of competent *E. coli* DH5α cells, 6 white colonies were selected and plasmids were isolated. Two of 6 clones had an expected length of insert. As a result of sequencing, it was found that one of the clones had a sequence whose deduced amino acid sequence was similar to known mevalonate pyrophosphate decarboxylase genes. This cDNA clone was designated as pMPD129 and used for further study.

[0092] Next, a partial genomic fragment which contained *mpd* gene was cloned by PCR. As a result of PCR whose condition was the same as that of the cloning of a partial cDNA fragment the amplified 1.05 kb fragment was obtained and was cloned into pMOSBlue T-vector. As a result of sequencing, it was confirmed that a genomic fragment containing *mpd* gene which had typical intron structures have been obtained and this genomic clone was named as pMPD220.

[0093] Southern blot hybridization study was performed to clone a genomic fragment which contained the entire *mpd* gene from *P. rhodozyma*. Probe was prepared by labeling a template DNA, pMPD220 digested by *KpnI*, with DIG multipriming method. Hybridization was performed with the method specified by the manufacturer. As a result, the probe hybridized to a band that had 7.5 kb in its lengths. Next, a genomic library consisting of from 6.5 to 9.0 kb of *EcoRI* fragment in the λZAPII vector was constructed. The packaged extract was infected to *E. coli* XL1Blue, MRF' strain and overlaid with NZY medium poured onto LB agar medium. About 6000 plaques were screened by using the 0.6 kb fragment of *KpnI*- digested pMPD220 as a probe. 4 plaques were hybridized to the labeled probe. Then a phage lysate was prepared according to the method specified by the manufacturer (Stratagene) and *in vivo* excision was performed by using *E. coli* XL1Blue MRF' and SOLR strains. Each 3 white colonies derived from 4 positive plaques were selected and plasmids were isolated from those selected transformants. Then, isolated plasmids were subjected to a colony PCR method whose protocol was the same as that in example 8. PCR primers whose sequences were shown in TABLE 14, depending on the sequence found in pMPD129 were synthesized and used for a colony PCR.

TABLE 15

Sequence of primers used in the colony PCR to clone a genomic *mpd* clone

Mpd7 ; CCGAACTCTCGCTCATCGCC (sense primer) (SEQ ID NO: 36)
--

TABLE 15 (continued)

Sequence of primers used in the colony PCR to clone a genomic *mpd* clone

Mpd8 ; CAGATCAGCGCGTGGAGTGA (antisense primer) (SEQ ID NO: 37)

[0094] PCR condition was almost the same as the cloning of *mvk* gene; 25 cycles of 30 seconds at 94 °C, 30 seconds at 50 °C and 10 seconds at 72 °C. All the clone except one produced a positive 0.2kb PCR band. A plasmid was prepared from one of the positive clones and the plasmid was named as pMPD701 and about 3 kb of sequence thereof was determined by the primer walking procedure (SEQ ID NO: 4). There existed an ORF consisted of 402 amino acids (SEQ ID NO: 9) whose sequence was similar to the sequences of known mevalonate pyrophosphate decarboxylase (52.3 % identity to mevalonate pyrophosphate decarboxylase from *Schizosaccharomyces pombe*). Also determined was a 0.4 kb of 5'-adjacent region which was expected to include its promoter sequence.

Example 13 Cloning of farnesyl pyrophosphate synthase (*fps*) gene

[0095] A cloning protocol of *fps* gene was almost the same as the *hmc* gene shown in Example 2 to 7. At first, the PCR primers whose sequence were shown in TABLE 16 based on the common sequences of farnesyl pyrophosphate synthase genes from other species were synthesized.

TABLE 16

Sequence of primers used in the cloning of *fps* gene

Fps1 ; CARGCNTAYTTYTNGTNGCNGAYGA (sense primer) (SEQ ID NO: 38)

Fps2 ; CAYTTRTTRTCYTGDATRTCNNGTNCDDATYTT (antisense primer) (SEQ ID NO: 39)

(N=A, C, G or T; R=A or G, Y=C or T, D=A, G or T)

[0096] After the PCR reaction of 25 cycles of 95 °C for 30 seconds, 54 °C for 30 seconds and 72°C for 30 seconds by using ExTaq as a DNA polymerase, a reaction mixture was applied to agarose gel electrophoresis. A PCR band that has a desired length (0.5 kb) was recovered and purified by QIAquick according to the method prepared by the manufacturer and then ligated to pUC57 vector. After a transformation of competent *E. coli* DH5α cells, 6 white colonies were selected and plasmids were then isolated. One of the plasmids which had desired length of an insert fragment was sequenced. As a result, it was found that this clone had a sequence whose deduced amino acid sequence was similar to known farnesyl pyrophosphate synthase genes. This cDNA clone was named as pFPS107 and used for further study.

[0097] Next, a genomic fragment was cloned by PCR by using the same primer set of Fps1 and Fps2. The same PCR condition as the case of cloning of a partial cDNA was used. A 1.0 kb band yielded was cloned and sequenced. This clone contained the same sequence with the pFPS107 and some typical intron fragments. This plasmid was named as pFPS113 and used for a further experiment.

[0098] Then, also cloned was a 5'- and 3'- adjacent region containing *fps* gene with the method described in Example 8. At first, the PCR primers whose sequences were shown in TABLE 17 were synthesized.

TABLE 17

Sequences of primers used for a cloning of adjacent region of *fps* gene

Fps7 ; ATCCTCATCCCGATGGGTGAATACT (sense for downstream cloning) (SEQ ID NO: 40)

Fps9 ; AGGAGCGGTCAACAGATCGATGAGC (antisense for upstream cloning) (SEQ ID NO: 41)

[0099] Amplified PCR bands were isolated and cloned into pMOSBlue T-vector. As a result of sequencing, it was found that the 5'-adjacent region that had 2.5 kb in its length and 3'-adjacent region that had 2.0 kb in its length were cloned. These plasmids were named as pFPSSTu117 and pFPSSTd117, respectively. After sequencing of both plasmids, it was found that an ORF that consisted of 1068 basepairs with 8 introns. Deduced amino acid sequence showed an extensive homology to the known farnesyl pyrophosphate synthase from other species. Based on the sequence determined, two PCR primers were synthesized with the sequences shown in TABLE 17 to clone a genomic *fps* clone and cDNA clone for *fps* gene expression in *E. coli*.

TABLE 18

Sequences of primers used for a cDNA and genomic *fps* cloning

Fps27 ; GAATTCATATGTCCACTACGCCTGA (sense primer) (SEQ ID NO: 42)

Fps28 ; GTCGACGGTACCTATCACTCCCGCC (antisense primer) (SEQ ID NO: 43)

[0100] PCR condition was as follows; 25 cycles of 30 seconds at 94 °C, 30 seconds at 50 °C and 30 seconds at 72 °C. One cDNA clone that had a correct sequence was selected as a result of sequencing analysis of clones obtained by PCR and was named as pFPS113. Next, Southern blot hybridization study was performed to clone a genomic fragment which contained the entire *fps* gene from *P. rhodozyma*. Probe was prepared by labeling a template DNA, pFPS113 with DIG multipriming method. As a result, labeled probe hybridized to a band that had about 10 kb in its length.

[0101] Next, a genomic library consisting of 9 to 15 kb of *EcoRI* fragment was constructed in a λ DASHII vector. The packaged extract was infected to *E. coli* XL1 Blue, MRA(P2) strain (Stratagene) and over-laid with NZY medium poured onto LB agar medium. About 10000 plaques were screened by using the 0.6 kb fragment of *SacI*- digested pFPS113 as a probe. Eight plaques were hybridized to the labeled probe. Then a phage lysate was prepared according to the method specified by the manufacturer (Promega). All the plaques were subjected to a plaque PCR using Fps27 and Fps28 primers. Template DNA for a plaque PCR was prepared by heating 2 μ l of solution of phage particles for 5 minutes at 99 °C prior to a PCR reaction. PCR condition is the same as that of pFPS113 cloning hereinbefore. All the plaques gave a 2 kb of positive PCR band, and this suggested that these clones had an entire region containing *fps* gene. One of the λ DNA that harbored *fps* gene was digested with *EcoRI* to isolate 10 kb of *EcoRI* fragment and to clone in *EcoRI*-digested and CIAP-treated pBluescriptII KS- (Stratagene). Twelve white colonies from transformed *E. coli* DH5 α cells were selected and plasmids were prepared from these clones and subjected to colony PCR by using the same primer sets of Fps27 and Fps28 and the same PCR condition. Two kb of positive band were yielded from 3 of 12 candidates. One clone was cloned and named as pFPS603. It was confirmed that sequence of *fps* gene which was previously determined from the sequence of pFPSSTu117 and pFPSStd117 were almost correct although they had some PCR errors. Finally, it was determined the nucleotide sequence of 4092 base pairs which contains *fps* gene from *P. rhodozyma* (Fig. 3), and an ORF which consisted of 365 amino acids with 8 introns was found (SEQ ID NO: 5). Deduced amino acid sequence (SEQ ID NO: 10) showed an extensive homology to known FPP synthase (65 % identity to FPP synthase from *Kluyveromyces lactis*).

SEQUENCE LISTING

(1) GENERAL INFORMATION:

- (i) APPLICANT: F.HOFFMANN-LA ROCHE. AG
- (ii) TITLE OF INVENTION: Improvement of microbiological carotenoid production and biological materials therefor
- (iii) NUMBER OF SEQUENCES: 43
- (iv) CORRESPONDENCE ADDRESS:
 - (A) ADDRESSEE:
 - (B) STREET: Grezacherstrasse 124
 - (C) CITY: BASLE
 - (E) COUNTRY: SWITZERLAND
 - (F) ZIP: CH-4002
- (v) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk
 - (B) COMPUTER: IBM PC compatible
 - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
- (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER:
 - (B) FILING DATE:
 - (C) CLASSIFICATION:
- (ix) TELECOMMUNICATION INFORMATION:
 - (A) TELEPHONE: 061-688 25 11
 - (B) TELEFAX: 061-688 13 95
 - (C) TELEX: 962292/965542 hlr c

(2) INFORMATION FOR SEQ ID NO:1:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 6370 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
 - (A) ORGANISM: Phaffia rhodozyma
 - (B) STRAIN: ATCC96594
- (ix) FEATURE:
 - (A) NAME/KEY: exon
 - (B) LOCATION: 1441..1466
- (ix) FEATURE:
 - (A) NAME/KEY: intron
 - (B) LOCATION: 1467..1722
- (ix) FEATURE:
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EP 0 955 363 A2

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 (A) NAME/KEY: intron
 (B) LOCATION: 1814..1914

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 (A) NAME/KEY: exon
 (B) LOCATION: 5524..5756

(ix) FEATURE:
 (A) NAME/KEY: polyA_site

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EP 0 955 363 A2

(B) LOCATION: 6173

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

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10	CGGGAGATCG TGGTTACATC AAGCCATCGT CACTGTAAGG CTCTGTAAGG CTCGGTTGTT	240
	AAGAAGGTAA CCAAGTGTA TCACTTGGTT CGCGGGGTGA CACTTAGGCT CTGGCGATTA	300
	ATATATCTGA AGCAGACCAA ACTATTAACA ATATACTTTT GGATAAGAGG TTTCAACAAG	360
15	AATCTCAGCT TGAGGAAAAC TCTTATCCAA GAAGGCGCGA GGGCGTCCCC GTTTTATATC	420
	AGGACCCCTC GCGCATTTGG TCTGCCACTA AAGATATACA TATGACGAGC CTAGAGAGGC	480
	TCGAGATCAC GAAAACTAAA AAGATGAAGC ATGAACCATG CAACTAGAG CATGATGGAA	540
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EP 0 955 363 A2

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10	AAGGACTTTA CCCTCGCAAA ATCTGTGTTT AGCTCCCCAG AAATCGCACC CGTCATGCTT	2220
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	AGAGTGGCCG CCAACAAGGT TGACATTGAG TGGGCCCCTC CGGTCGCCG CTCCCGTATC	2340
15	GTCATTGGCT CGGTGAGAA GATCGGGTCC TCGATCGTCA GAGACTTTGC CCTCGAGGTC	2400
	CCCCTCTCC TTCTCGGAGC CGCCAGCGGG CTCGGCGGAC TCAAGGAGTT TTGTAAGCTC	2460
	GCCGCGTTAA TTTTGGTGGC CGACTGCTGC TTCACCTTTA CCTTCTATGT CGCCATCCTC	2520
20	ACCGTCATGG TCGAGGTAAG CCTTTCTTC AAGTTTCTTG CTGTCATTTT CTTTTCGACA	2580
	CGTATGCTCA TCTTTCGTTT CCGTCTCTCT CACCTTTCCA GGTCACCGA ATCAAGATCA	2640
	TCCGGGGCTT CCGACCGGCC CACAATAACC GAACACCGAA TACTGTGCCC TCTACCCCTA	2700
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35	TTAGTGACTG GACGACCATC GTCGGAGATC CAATCATGAG CAAGTGGATC ATCATCACC	3180
	TGGGCGTGT CATCTGCTG AACGGGTTC TCCTAAAAGG GATCGCTTCT GGCTCTGCTC	3240
	TGCGACCCGG TCGTCCCGGA GGAGGAGGAG CTGCCGCCG CGCCGCCGTC TTGCTCGGAG	3300
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EP 0 955 363 A2

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 30 TCTTTCATA TTCGTCTCTG TTTAATTTCT TTTCTGTCCA GTCTTATGAC GTCTGATTGG 4980
 TTCTTCTTTT CACCCACACA CATAAGTCA ATCTTCTTGG CTACAGGTCA GGATCCTGCA 5040
 CAGAATGTGG AGTCTTCAAT GTGCATGACA TTGATGGAGG CGTACGTTTT TTGTTTTGTT 5100
 35 TTCTTCTTTT TTCCATATGT TTCTACTTCT ACTTCTTCC CGAGTCCGCC AAGCTGATAC 5160
 CTTTATACGG TCCTTCTCTT TCTCATGACG AGTAGTGTGA ACGACGGAAA AGATCTACTC 5220
 ATCACCTGCT CGATGCCGGC GATCGAGTGC GGAACGGTCG GTGGAGGAAC TTTCTTCCCT 5280
 40 CCGCAAAACG CCGTTTGTGA GATGCTCGGT GTCGCAGGTG CCCATCCAGA TTCGCCCGGT 5340
 CACAATGCTC GTCGACTAGC AAGAATCATC GCTGCCAGTG TGATGGCTGG AGAGTTGAGT 5400
 TTGATGAGTG CTTTGGCCGC TGGTCATTTA ATCAAGGCCC ACATGAGTAA GTCTGCCACC 5460
 TTTTGATAAT CAAAAGGGTC GTGGTACTGG TGTCACTGAC TGGTGAATCT TCCTGTCTATG 5520
 45 CAGAGCACAA TCGATCGACA CCTTCGACTC CTCTACCGGT CTCACCGTTG GCGACCCGAC 5580
 CGAACACGCC GTCCACCGG TCGATTGGAT TGCTCACACC GATGACGTCT TCCGCATCGG 5640
 TCGCCTCGAT GTTCTCTGGG TTCGGTAGTC CGTCGACGAG CTCGCTCAAG ACGGTAGGTA 5700
 50 GCATGGCTTG CGTCAGGGAA CGAGGGGACG AGACGAGTGT GAACGTGGAT GCCTGAACTG 5760
 GGGACTCCCT TTTCTTGGTA TCCCTTCCGT TTTTCTTTTC GCCTTTGAAT CCTGTATTCT 5820

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TGTCCGTTTT TTCATCTTCT CTTCCTGGTT CTCCTTCTCT CGTTCATCTG CAAAAACAAA 5880
 ATTCAATCGC ATCGGTCTCT GGCATTCAT TTGGGTTTCA AAATCAAATC AATCTCTATC 5940
 5 TACTATCTCA AATATCTTTT TTTCATCTTT TGATTCATTT CTGTTGAAAA CTGTCCTGCC 6000
 CTCTCTCTAC TTCTTATCTC TGCCTTCTTG CCAAAGTTCA ATTCGTTGTC CATCTGTGCA 6060
 CTCTGATCTA TCAGTCGTGA TCAAGTACGC TCTTAAATCT GTAATTGGCT CTCGGAGGTG 6120
 10 TCTCGTCATC TCACATATGG CTGGCGATAT GATGTGTCGG TTTCTTCCCC TCCAACAAAG 6180
 GCGACGTGGC TCCTTCATCA ATCTTTGGCG CAAGCTCTCA AAATCTCCA AAACGGCTGA 6240
 CTAAGCAAGG TTTCCAAGTA CTCTCAAACC GAGCAAGGCC ATCCATCCTC AAATCAACTT 6300
 15 GTGAAACCCCT TTGTGGATAG ACCGTCCAAA CCGAGCTCTT CCCAATCTTC GCCTCCCTT 6360
 CTTCCTGCAG 6370

(2) INFORMATION FOR SEQ ID NO:2:

20 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 4775 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear
 25 (ii) MOLECULE TYPE: DNA (genomic)
 (iii) HYPOTHETICAL: NO
 (iv) ANTI-SENSE: NO
 (vi) ORIGINAL SOURCE:
 30 (A) ORGANISM: *Phaffia rhodozyma*
 (B) STRAIN: ATCC96594
 (ix) FEATURE:
 (A) NAME/KEY: exon
 (B) LOCATION: 1305..1361
 35 (ix) FEATURE:
 (A) NAME/KEY: intron
 (B) LOCATION: 1362..1504
 (ix) FEATURE:
 40 (A) NAME/KEY: exon
 (B) LOCATION: 1505..1522
 (ix) FEATURE:
 (A) NAME/KEY: intron
 (B) LOCATION: 1523..1699
 45 (ix) FEATURE:
 (A) NAME/KEY: exon
 (B) LOCATION: 1700..1826
 (ix) FEATURE:
 (A) NAME/KEY: intron
 (B) LOCATION: 1827..1920
 50 (ix) FEATURE:
 (A) NAME/KEY: exon
 (B) LOCATION: 1921..2277
 55

(ix) FEATURE:
 (A) NAME/KEY: intron
 (B) LOCATION: 2278..2351

5

(ix) FEATURE:
 (A) NAME/KEY: exon
 (B) LOCATION: 2352..2409

(ix) FEATURE:
 (A) NAME/KEY: intron
 (B) LOCATION: 2410..2497

10

(ix) FEATURE:
 (A) NAME/KEY: exon
 (B) LOCATION: 2498..2504

15

(ix) FEATURE:
 (A) NAME/KEY: intron
 (B) LOCATION: 2505..2586

(ix) FEATURE:
 (A) NAME/KEY: exon
 (B) LOCATION: 2587..2768

20

(ix) FEATURE:
 (A) NAME/KEY: intron
 (B) LOCATION: 2769..2851

(ix) FEATURE:
 (A) NAME/KEY: exon
 (B) LOCATION: 2852..2891

25

(ix) FEATURE:
 (A) NAME/KEY: intron
 (B) LOCATION: 2892..2985

30

(ix) FEATURE:
 (A) NAME/KEY: exon
 (B) LOCATION: 2986..3240

(ix) FEATURE:
 (A) NAME/KEY: intron
 (B) LOCATION: 3241..3325

35

(ix) FEATURE:
 (A) NAME/KEY: exon
 (B) LOCATION: 3326..3493

(ix) FEATURE:
 (A) NAME/KEY: intron
 (B) LOCATION: 3494..3601

40

(ix) FEATURE:
 (A) NAME/KEY: exon
 (B) LOCATION: 3602..3768

45

(ix) FEATURE:
 (A) NAME/KEY: polyA_site
 (B) LOCATION: 4043

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

50

CATCGAAGAG AGCGAAGTGA TTAGGGAAGC CGAAGAGGCA CTAACAACGT GGTGTATAT
GTGTGTTTAT GAGTGTATA TCGTCAAGAA CGAAGTCCAT TCATTTAGCT AGACAGGGAG

60

120

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EP 0 955 363 A2

	AGAGGGAGAA ACGTACGGGT TTACCCCTATT GGACCAGTCT AAAGAGAGAA CGAGAGTTT	180
5	TGGGTCGGTC ACCTGAAGAG TTTGAACCTC CACAAGTTTA TTCTAGATTA TTCCGGGGG	240
	TATGTGAAGG ATAATGTCAA ACTTTGTCCA GATTGAAGAA GGCAAGAAAG GAAAGGGGCG	300
	AACGAGAGTA TCGTCCCATC TATGGGTGAC CAGTCGACCT TCTGCATCGG CGATCCCGAG	360
	AATGGAAGGT TCCGATGGAT CAGAAGTAGG TTTCTAAGC TCAAACATAG GTCATTGCGA	420
10	GTGAGATACA TATGCAGACT GATATGCTAG TCAAACCGAA CGAGATTCT CTGTTTGCTT	480
	TCAAAAAGAC GAACCAACCA TTTCATGTCC AAGATGGCAG GTCTTTCGAT TCTTTGAAGC	540
	TCCTCCCTGA TCCGGACAGA AAAGAATAAA AAGTAGACAG ACTGTCAAGT CGACAGCGCA	600
15	AGTTTATCAA GCTGAGCGAG AAAACTCGAA CTTACATACC TTGGCCGTC GTTCTGTAGA	660
	CCAAGCATCG GCCPTTCTC TTTGCGGCAG GTGTACCGT TGGCTCACCA TCGTCACTCT	720
	CGTCTCCTGA CCCGTTGCTT TCCTTGACAG CAGTCTGTT CACAGGTTT TCTAACTGAT	780
20	AGGTCCCAAC AGCAAAGATA TCTGGATGTC TATGTGAGAA CTCTACTGAG TCGGCAGAGT	840
	ACACCGTATC GATATAGGCG AGTGAGGAAG CTTTGAAAGG TGAAGAAGTA GCGAAAGATC	900
	ATCAGCGAAT GAGGACTATG ACAAAAAAGA AATTTTCGTA TAATCCACTG GACAAATCAC	960
25	CTTCCATCGT GTCCTCCAAG AGGGTTTCGT CTGAAACGTA AGGACGAGGT ATTGATAGAT	1020
	GATTGACCTT GAGTACGCGG ATGGACAAGG AACGAGCCCA CTCCAGGGC TATGTAACAC	1080
	CACACGTGAC TCCACTTGAA TTGCGGCAGA TAAACGAAGT CTTACGATCG GACGACTTTG	1140
	TAACCATTTA GTTATTTACC CGTCTTGTTT TCTTACTTTG ATCGTCCCAT TTTAGACACA	1200
30	AAAAAAGAAG CCAGAAGAGA AAAGAATAAA ACGTCTACCG TGTCTCTCC GAATTCTTAC	1260
	CACACCACA AAACCATAACA CAATCTCAAT CTAGATATCC AGTTATGTAC ACTTCTACTA	1320
	CCGAACAGCG ACCCAAAGAT GTTGAATTC TCGGTATGGA GGTATGTTGT TCAATCTCTG	1380
35	TTGTGTCAA TCTTTAATCA TCTTTAGTCG ACTGACCGGT TCTTCCTTT TTTTCTTCA	1440
	TCAAACAAA CAACCTTCT CGATTCTATG CATCTTTCTT TCCAATGCGC TACTCCTTCT	1500
	GTAGATCTAC TTTCTCGAC GAGTGGCTAA CTATTCTCTC TTCTGCATTG TCTCTCTATT	1560
40	CCCATGTTG ATCCCTCGCC CTCATATGGG CGACTGTTT ATCTCTTTT CTTCCGTCCA	1620
	TTCTTCTTG ATCTGTTC TTTTCTACTA ATATCTCCCG ACCCGAAATA CAACACTGAC	1680
	CGCGATTCT CTCGATCAGG CCATCGCTCA CAAGGATCTC GAGGCTTTTG ATGGGGTTCC	1740
45	TTCCGGAAG TACACCATCG GTCTCGGCAA CAACTTCATG GCCTTCACCG ACGACACTGA	1800
	GGACATCAAC TCGTTCGCCT TGAACGGTCA GTCTCTTCCG TTTCAGCAAT CGACAGGAAA	1860
	AAGGCCAAG CGCATCTCAC TGACACCTT CTCCGTTTG CAATTCCATT TGATTGTTAG	1920
	CTGTTTCCGG TCTTCTATCA AAGTACAACG TTGATCCCAA GTCAATCGGT CGAATTGATG	1980
50	TCGGAACCTA GTCCATCATT GACAAGTCCA AATCTGTCAA GACAGTCTT ATGGACTTGT	2040
	TCGAGTCCCA CGGCAACACA GATATTGAGG GTATCGACTC CAAGAATGCC TGCTACGGTT	2100
55		

EP 0 955 363 A2

	CTACCGCGGC CCTGTCAAT GCCGTCAACT GGATCGAGTC ATCCTCTTGG GACGGAAGAA	2160
5	ATGCCATTGT CTTCTGCGGA GACATTGCCA TCTACGCCGA GGGTGCTGCC CGACCTGCCG	2220
	GAGGTGCTGG TGCTTGCGCC ATCCTCATCG GACCCGACGC TCCCGTCGTC TTCGAGCGTG	2280
	AGTTCCAATC CGTCATTTTC TTCCACGGCA GCGGCTGAAA CAACCCCTAT CCGTCATTCT	2340
	CATCAATCTA GCCGTCCACG GAAACTTCAT GACCAACGCT TGGGACTTCT ACAAGCCTAA	2400
10	TCTTTCTTCG TATGTTCAAA TTTTGAAGTT TGGGCTTGGG AGAGTCTTAC ACTAATTCGG	2460
	GGTGCTCGTA TCCTTCGAAT CGTTTGTTC TTTATAGTGA ATACGTCGT CTGCGCACCT	2520
	CCTATATTTA GTTTTGTATC AAATATTGTC CATTGAATTA ACTCTGAAAC CTTCTCCTCC	2580
15	AAATAGCCCA TTGTCGATGG ACCTCTCTCC GTCACCTCCT ACGTCAACGC CATTGACAAG	2640
	GCCTATGAAG CTTACCGAAC AAAGTATGCC AAGCGATTG GAGGACCCAA GACTAACGGT	2700
	GTCACCAACG GACACACCGA GGTGCGCGT GTCAGTGCTG CGTCGTCGA TTACCTTTTG	2760
20	TTCCACAGGT AAGCGTCATC TTCTGTATTC TCCTTAAAT CAACCGATCA ACGGAGTTAA	2820
	TTCGTGTCAT CATATTATCT TGTGGAACA GTCCTTACGG AAAGCAGGT GTCAAAGGCC	2880
	ACGGCCGACT TGTAAGCAGT CTTTTGTAA CTCTTAGCTT GCAGATAAAA ACTTTTAGGT	2940
25	TTCTGGTACT CATTATTTAT GCATCTCTTG AATCACCTTA TCTAGTTGTA CAATGACTTC	3000
	CGAAACAACC CCAACGACCC GGTTTTGTCT GAGGTGCCAG CCGAGCTTGC TACTTTGGAC	3060
	ATGAAGAAAA GTCTTTCAGA CAAGAATGTC GAGAAATCTC TGATTGCTGC CTCCAAGTCT	3120
	TCTTTCAACA AGCAGGTGTA GCCTGGAATG ACCACCGTCC GACAGCTCGG AAACCTGTAC	3180
30	ACCGCCTCTC TCTCGGTGC TCTCGCAAGT TTGTTCTCTA ATGTTCTTGG TGACGAGCTC	3240
	GTAAGTCTTG ATCTCTATCC CAATCATCTC TTCCTTATCA ATTGAACTGA ACTCTTTTCT	3300
	TTAATGCTGG CTTTCTCTTG AACAGGTCGG CAAGCGCATT GCTCTCTACG CCTACGGATC	3360
35	TGGAGCTGCT GCTTCTTTCT ATGCTCTTAA GGTCAAGAGC TCAACCGCTT TCATCTCTGA	3420
	GAAGCTTGAT CTCAACAACC GATTGAGCAA CATGAAGATT GTCCCCTGTG ATGACTTTGT	3480
	CAAAGCTCTG AAGGTACGTT GGATAATGAC TTTTITGTG GACCGTGGTC TTTGTCAACC	3540
40	GCTAACAACC TTCTTGAATC GTCTCTTTT GGTITGAAAT TCGCTCGGCG CTTGACACA	3600
	GGTCCGAGAA GAGACTCACA ACGCCGTGTC ATATTGCCCC ATCGGTTTCG TTGACGATCT	3660
	CTGGCCTGGA TCGTACTACT TGGGAGAGAT TGACAGCATG TGGCGTCGAC AGTACAAGCA	3720
45	GGTCCCTTCT GCTTGAACGG GATATTAAAA GTTTCAAAAG TTATGAAAGA GGTCCGCGAA	3780
	GATTCAAAAT AAATAAATAT AACACCTTGC TTTTGGCTT GTTTTCCCTC TTCACTCTCG	3840
	TTTCCGATGT GTTCTCTCCG TTTCTTCCCT CTTTGTTCCT TTTTCTCTCC CTCTTTTGGT	3900
	TACAATCTCT TTGGGTTTTA CAGGCTGGCA ATCTCTGTAC AATCTTCGTT CGCGTGATCC	3960
50	GACATAGATA CCGTTGTGGC ATACACCTTG CGTCTTACAT CTTTGTAGAG CTTGCGAGGT	4020
	GATCTTGATG AAGAAAATTC ACCATTGACT CCCATCTCTT GAATGTCCTG ACTAAATTGA	4080
55		

ATTGGAAGCA ACTTATATGA AGAGCAAATT GATGGATCCA GAAAGGAACA AGTCTAGAAA 4140
 TCAGTGATTT GTGCGAAAAA TCAGCAAATG CCGCGCTGAG CCGCTCGCTG GGGAGTAGAC 4200
 5 ATTGCCCATG CGCGTGATGT TGTCTGACCG TTCTCCTCCA TTCCCCCACT CTCAACCTTC 4260
 CTCTCTTTGA GAATCGAAGA AGAAGGCGAA GAAAACCTGA CTTGATCCTT TACAGGGTGT 4320
 TTCTTTTGTT CGTATCTGAG TTACTTTTCC TCCTTTCCCTT CCTGCTTGAG TGAATGACTG 4380
 10 ATCTGACTCC TCCGCCTACC TCGGCGACTG GGCTATATCT TGAGGATAGA ATATCCCCCT 4440
 GACAATCCCA TTCTCAAGA TTCTTTCAAA CAAGAAACT AGTTCCAATC AATAGATCAT 4500
 CTGATCAACC TTGTGTGAAC ATAATCATCT GCAGAAGCAC TGAAGTGAAGA AAGTCTTCCT 4560
 15 CAGAGGAAAG AGAATACTAG ATAAGATCAT TCGGTTGGGA AGGTAAAGGA ATGAAGTCTG 4620
 GTTCTGGGTT TAGCTCTGGT TCCGTAGGGG GTTCGACTAT AGTTTCTTCT GTTCGACTAG 4680
 AAACAGGAGA AACCGTACAT GTAAATGGTA TGATATTCTT GTCTCTGTAT CATGTCCCGC 4740
 20 TCATCTCTTT GTTGCAAGT CACTCTGGAG AATTC 4775

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 4135 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: *Phaffia rhodozyma*
 (B) STRAIN: ATCC96594

(ix) FEATURE:

- (A) NAME/KEY: exon
 (B) LOCATION: 1021..1124

(ix) FEATURE:

- (A) NAME/KEY: intron
 (B) LOCATION: 1125..1630

(ix) FEATURE:

- (A) NAME/KEY: exon
 (B) LOCATION: 1631..1956

(ix) FEATURE:

- (A) NAME/KEY: intron
 (B) LOCATION: 1957..2051

(ix) FEATURE:

- (A) NAME/KEY: exon
 (B) LOCATION: 2052..2366

(ix) FEATURE:

- (A) NAME/KEY: intron

EP 0 955 363 A2

(B) LOCATION: 2367..2446

(ix) FEATURE:

(A) NAME/KEY: exon

(B) LOCATION: 2447..2651

(ix) FEATURE:

(A) NAME/KEY: intron

(B) LOCATION: 2652..2732

(ix) FEATURE:

(A) NAME/KEY: exon

(B) LOCATION: 2733..3188

(ix) FEATURE:

(A) NAME/KEY: polyA_site

(B) LOCATION: 3284

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

ACTGACTCGG CTACCGGAAA ATATCTTTTC AGGACGCCTT GATCGTTTTG GACAACACCA	60
TGATGTCACC ATATCTTTCAG CGGCCGTTGG AGCTAGGAGT AGACATTGTA TACGACTCTG	120
GAACAAAGTA TTTGAGTGGA CACCACGATC TCATGGCTGG TGTGATTACT ACTCGTACTG	180
AGGAGATTGG GAAGGTTTCGT GCTTGCTTGC TTTGAATGTC GTGCCCTAAAG CCATTGCCAT	240
AAGACAGAGT CTGATCTATG TCGTTTGCCT ACAACAGAGA ATGGCCTGGT TCCCAAATGC	300
TATGGGAAAT GCATTGTCTC CGTTCGACTC GTTCCTTCTT CTCCGAGGAC TCAAAACACT	360
TCCTCTCCGA CTGGACAAGC AGCAGGCCCTC ATCTCACCTG ATCGCCTCGT ACTTACACAC	420
CCTCGGCTTT CTGTTCCTACT ACCCCGGTCT GCCTTCTGAC CCTGGGTACG AACTTCATAA	480
CTCTCAGGCG AGTGGTGCGAG GTGCCGTTCAT GAGCTTTGAG ACCGGAGATA TCGCGTTGAG	540
TGAGGCCATC GTGGGCGGAA CCCGAGTTTG GGAATCAGT GTCAGTTTCG GAGCCGTGAA	600
CAGTTTGATC AGCATGCCTT GTCTAATGAG GTTAGTTCTT ATGCCCTTCTT TTCGCGCCTT	660
CTAAAATTTT TGGCTGACTA ATTGGGTCGG TCTTTCCGTT CTTCGATTTT AGTCACGCAT	720
CTATTCCTGC TCACCTTCGA GCCGAGCGAG GTCTCCCCGA ACATCTGATT CGACTGTGTG	780
TCGGTATTGA GGACCCTCAC GATTTGCTTG ATGATTGGA GGCTCTCTT GTGAACGCTG	840
GCGCAATCCG ATCAGTCTCT ACCTCAGATT CATCCCGACC GCTCACTCCT CCTGCCTCTG	900
ATTCTGCCTC GGACATTCAC TCCAACTGGG CCGTCGACCG AGCCAGACAG TTCGAGCGTG	960
TTAGGCCTTC TAACTCGACA GCCGGCGTCG AAGGACAGCT TGCCGAACCTC AATGTAGACG	1020
ATGCAGCCAG ACTTGCGGGC GATGAGAGCC AAAAAGAAGA AATTCCTTGT AGTGACCCGG	1080
GAAAGGTCAT TCTGTTCCGC GAACATGCTG TAGGCCATGG TGTGTGAGT GAGAAATGAA	1140
AGCTTTATGC TCTCATTGCA TCTTAACCTT TCCTCGCCCT TTTTGTCTC TTCATCCCGT	1200
CTTGATTGTA GGGATGCCCC CCTTTGCCCTC TTTCCCTTC TTGCATCTGT CTATATTTCC	1260
TTATACATTT CGCTCTTAAG AGCGTCTAGT TGTACCTTAT AACAACCTTT GGTTTTAGCA	1320
TCCTTTGATT ATTCATTTCT CTCATCCTTC GGTGAGAGGC TTTCGGCCAT CTTTACGTCT	1380

EP 0 955 363 A2

	GATTAGATTG TAATAGCAAG AACTATCTTG CTAAGCCTTT TCTCTTCCTC TTCCTCCTAT	1440
	ATAAATCGAA TTCACTTTTCG GACATGTTTA TTTTGGGGAA ATCATCAAGG GGTGGGGGGC	1500
5	CAATCCCGAC ACTAATTTTC TGCTCACGTC AAAACTCAGC GTTCAGAATC AGTCACTGAC	1560
	CCTGATACGT GTCTCTATGT GTGTGGGTGT ACGTGCGAAT TGTGACTCGA CGTCTACGC	1620
	TTAAAAACAG ACCGGGATCG CTGCTTCCGT TGATCTTCGA TGCTACGCTC TTCTCTCACC	1680
10	CACTGCTACG ACAACAACAT CATCGTCGTT ATCGTCTACA AACATTACCA TCTCCCTAAC	1740
	GGACCTGAAC TTTACGCAGT CTTGGCCTGT TGATTCTCTT CCTTGGTCAC TTGCGCCTGA	1800
	CTGGACTGAG GCGTCTATTC CAGAATCTCT CTGCCCCACA TTGCTCGCCG AAATCGAAAG	1860
15	GATCGCTGGT CAAGGTGGAA ACGGAGGAGA AAGGGAGAAG GTGGCAACCA TGGCATTCTT	1920
	GTATTTGTTG GTGCTATTGA GCAAAGGGAA GCCAAGGTAG GTTTTTTCTG TCTCTCTTTT	1980
	TTGCCTATAA AGACTCTTAA CTGACGGAGA AAGTGTGGG TTTCTTCCTT CGGGGGTTCA	2040
20	ATCAATTAAA GTGAGCCGTT CGAGTTGACG GCTCGATCTG CGCTTCCGAT GGGAGCTGGT	2100
	CTGGGTTTCT CCGCCGCTCT ATCGACCTCT CTTGCCCTAG TCTTTCTTCT CCACTTTTCT	2160
	CACCTCAGTC CAACGACGAC TGGCAGAGAA TCAACAATCC CGACGGCCGA CACAGAAGTA	2220
25	ATTGACAAAT GGGCGTTCTT AGCTGAAAAA GTCATCCATG GAAATCCGAG TGGGATTGAT	2280
	AACGCGGTCA GTACGAGAGG AGGCGCTGTT GCTTTCAAAA GAAAGATTGA GGGAAAACAG	2340
	GAAGGTGGAA TGGAAGCGAT CAAGAGGTAC GCAGACACGG TGCTTCATAT GCCATACTCC	2400
	AGTCTGATTG ACCCATGATG AACGTCCTTC TACATTTTGA ATATAGCTTC ACATCCATTC	2460
30	GATTCCTCAT CACAGATTCT CGTATCGGAA GGGATACAAG ATCTCTCGTT GCAGGAGTGA	2520
	ATGCTCGACT GATTCAAGGAG CCAGAGGTGA TCGTCCCTTT GTTGAAGCG ATTCAAGAGA	2580
	TTGCCGATGA GGCTATTGCA TGCTTGAAAG ATTCAGAGAT GGAACGTGCT GTCATGATCG	2640
35	ATCGACTTCA AGTTAGTTCT TGTTCCTTTC AAGACTCTTT GTGACATTGT GTCTTATCCA	2700
	TTTCATCTTC TTTTTCCTTC CTCTCTCTGC AGAACTTGGT CTCCGAGAAC CACGCACACC	2760
	TAGCAGCACT TGGCGTGTCC CACCCATCCC TCGAAGAGAT TATCCGATC GGTGCTGATA	2820
40	AGCCTTTCGA GCTTCGAACA AAGTTGACAG GCGCCGCTCG AGGTGGTTGC GCTGTAACCC	2880
	TGGTGCCCGA TGGTAAAGTC TCTCCTTTTC TCTTCCGTCC AAGCGACACA TCTGACCGAT	2940
	GCGCATCCTG TACTTTTGGT CAACCAGACT TCTCGACTGA AACCCTTCAA GCTCTTATGG	3000
45	AGACGCTCGT TCAATCATCG TTCGCCCTTT ATATTGCCCG AGTGGGTGGT TCAGGCGTCG	3060
	GATTCCTTTC ATCAACTAAG GCCGATCCGG AAGATGGGGA GAACAGACTT AAAGATGGGC	3120
	TGGTGGGAAC GGAGATTGAT GAGCTAGACA GATGGGCTTT GAAAACGGGT CGTTGGTCTT	3180
	TTGCTTGAAC GAAAGATAGG AAACGGTGAT TAGGGTACAG ATCCTTTGCT GTCATTTTAA	3240
50	CAAAACACTT TCTTATGTCT TCATGACTCA ACGTATGCCC TCATCTCTAT CCATAGACAG	3300
	CACGGTACCT CTCAGTTTTC AATACGTAAG CGTTCATCGA CAAAACATGC GGCACACGAA	3360

EP 0 955 363 A2

AACGAGTGGG TATAAGGGAG AAGAGAGATA TTAGAGCGAA AAAGAGAAGA GTGAGAGAGG 3420
 AAAAAAATAA CCGAGAACAA CTTATTCCGG TTTGTTAGAA TCGAAGATCG AGAAATATGA 3480
 5 AGTACATAGT ATAAAGTAAA GAAGAGAGGT TTACCTCAGA GGTGTGTACG AAGGTGAGGA 3540
 CAGGTAAGAG GAATAATTGA CTATCGAAAA AAGAGAACTC AACAGAAGCA CTGGGATAAA 3600
 GCCTAGAATG TAAGTCTCAT CGGTCCGCGA TGAAAGAGAA ATTGAAGGAA GAAAAAGCCC 3660
 10 CCAGTAAACA ATCCAACCAA CCTCTTGAC GATTGCGAAA CACACACACG CACGCGGACA 3720
 TATTTCTGAC ACAAGGACGG GACATTCCTT TTTTATATCC GGGTGGGGAG AGAGAGGGTT 3780
 ATAGAGGATG AATAGCAAGG TTGATGTTTT GTAAAAGGTT GCAGAAAAG GAAAGTGAGA 3840
 GTAGGAACAT GCATTAAAAA CTGCCCCAAA GCGATTATA TCGTCTTCT GTTTTCACTT 3900
 15 CTTTCCGGGC GCTTCTTAG ACCGCGGTGG TGAAGGGTTA CTCCTGCCAA CTAGAAGAAG 3960
 CAACATGAGT CAAGGATTAG ATCATCACGT GTCTCATTG ACGGGTTGAA AGATATATTT 4020
 AGATACTAAC TGCTTCCAC GCCGACTGAA AAGATGAATT GAATCATGTC GAGTGGCAAC 4080
 20 GAACGAAAGA ACAAATAGTA AGAATGAATT ACTAGAAAAG ACAGAATGAC TAGAA 4135

(2) INFORMATION FOR SEQ ID NO:4:

25 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 2767 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

 (ii) MOLECULE TYPE: DNA (genomic)
 30 (iii) HYPOTHETICAL: NO
 (iv) ANTI-SENSE: NO
 (vi) ORIGINAL SOURCE:
 (A) ORGANISM: *Phaffia rhodozyma*
 35 (B) STRAIN: ATCC96594

 (ix) FEATURE:
 (A) NAME/KEY: exon
 (B) LOCATION: 401..451

 (ix) FEATURE:
 40 (A) NAME/KEY: intron
 (B) LOCATION: 452..633

 (ix) FEATURE:
 (A) NAME/KEY: exon
 (B) LOCATION: 634..876
 45 (ix) FEATURE:
 (A) NAME/KEY: intron
 (B) LOCATION: 877..1004

 (ix) FEATURE:
 50 (A) NAME/KEY: exon
 (B) LOCATION: 1005..1916

 (ix) FEATURE:

55

EP 0 955 363 A2

(A) NAME/KEY: polyA_site
(B) LOCATION: 2217

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

5	GAATTCCTCC CGACTGGGCT GATCGACTTG ACTGGAAGAT CTAAGGCGGA GGGATGAAGG	60
	AAGTAATTGG AGGGAATGAG GAAAAAAAAA GGCAGGGGAA CGCGTCTTC TTCTCTGGCA	120
	AGGCAATGTC GTGTATCTCT CTTGATTCTT TCGTTGTATC GACGGACCAC ACTCTTTTCG	180
10	AATGAATATC ACTATCGCAT CCAATGATCG CTATACATGG CATTTACATA TGCCAGACAT	240
	CGCTGAGAAA GAGAGAACAT TCCTTTGGAA AAAGCCTACT GTGCCTGAAG TCAGGCTGAT	300
	GTTGATTAAA CGTCTTTCCC CATCCTAAGC AGACAAACAA CTCTTTTTCG TTCAACACAC	360
15	CACCTCTCTC CGAAAAAGCT CTTCAATCCA GTCCATTAAAG ATGGTTCATA TCGCTACTGC	420
	CTCGGCTCCC GTTAACATTG CGTGTATCAA GGTCCGCTCG CATTTGTGAAT GCTGCTCGTT	480
	TGCCTTGTGT GCGTTTGGTG GATCTGAAAG AACCTTGTCT TGAACCATTC CATCTCTGCT	540
20	CTTTTCTTTC CTGTCTTTTC CTTTCTCTCA CGACAAAAAA ACCACCTGGA CCCTTTGTGT	600
	TCCTTTCCAT TGGTGTTCAT ACACCTAACA CAGTACTGGG GTAAACGGGA TACCAAGTTG	660
	ATTCTCCCTA CAAACTCCTC CTTGTCTGTC ACTCTCGACC AGGATCACCT CCGATCGACG	720
25	ACGTCTTCTG CTTGTGACGC CTCGTTTCGAG AAGGATCGAC TTGGGCTTAA CGGGATCGAG	780
	GAGGAGGTCA AGGCTGGTGG TCGGTTGGAT GTCTGCATCA AGGAGATGAA GAAGCTTCGA	840
	GCGCAAGAGG AAGAGAAGGA TGCCGGTCTG GAGAAAGTGA GTTTTCTTCC TGTGTGCGTG	900
	TGTACTCTGT ATAGGTACCG TTGACAGGAC AGTCTTCTG AAGAGTTTGG ATCTTACTCT	960
30	TTTTTGGGGG GGTGGTGGTG TTTGAAATAA TGACCAAAAT AAAGCTCTCA TCTTTCAACG	1020
	TGCACCTTGC GTCTTACAAC AACTTCCCGA CTGCCGCTGG ACTTGCTTCC TCCGCTTCCG	1080
	GTCTAGCTGC GTTGGTCGCC TCGCTCGCCT CGCTCTACAA CCTCCCAACG AACGCATCCG	1140
35	AACTCTCGCT CATCGCCCGA CAAGGTTCTG GTTCTGCCTG CCGATCGCTC TTCGGCGGGT	1200
	TCGTTGCTTG GGAACAGGGC AAGCTTTCCT CTGGAACCGA CTCGTTTCGT GTTCAGGTCG	1260
	AGCCCAGGGA ACACTGGCCC TCACTCCACG CGCTGATCTG TGTAGTTTCC GACGAGAAAA	1320
40	AGACGACGGC CTCGACGCCA GGCATGCAAA CCACGGTGAA CACCTCGCCT TTGCTCCAAC	1380
	ACCGAATCGA ACACGTCGTT CCAGCCCGGA TGGAGGCCAT CACCCAGGCG ATCCGGGCCA	1440
	AGGATTTCGA CTCGTTCCGA AAGATCACCA TGAAGGACTC CAACCAGTTC CACGCCGTCT	1500
45	GCCTCGATTG GGAACCCCGG ATCTTTTACT TGAACGATGT CTCCCGATCG ATCATCCATC	1560
	TCGTACCCGA GCTCAACAGA GTGTCCGTCC AGGCCGGCGG TCCCGTCCTT GCCGCCTACA	1620
	CGTTGACGCG CGGGCCGAAC GCGGTGATCT ACGCCGAGGA ATCGTCCATG CCGGAGATCA	1680
	TCAGGTTAAT CGAGCGGTAC TTCCCGTTGG GAACGGCTTT CGAGAACCCG TTCGGGGTTA	1740
50	ACACCGAAGG CGGTGATGCC CTGAGGGAAG GCTTTAACCA GAACGTCGCC CCGGTGTTCA	1800
	GGAAGGGAAG CGTCGCCCGG TTGATTACCA CCCGGATCGG TGATGGACCC AGGACGTATG	1860
55		

5 GCGAGGAGGA GAGCCTGATC GGCGAAGACG GTCTGCCAAA GGTCGTCAAG GCTTAGACTA 1920
 TAGGTTGTTT CTTCTAAATT TGAGCCTTCC TCCCGCCTCC CTTCCACAAG CATAAAACAA 1980
 AGGATAAACA AATGAATTAT CAAAATAACT ATAGGTTGTT TCTTCTAAAT TTGAGCCTTC 2040
 CTCCCGCCTC CCTTCCACAA GCATAAAACA AAGGATAAAC AAATGAATTA TCAAAATAAA 2100
 ATAAAAAGTC TGCCTTCTTT GTTTTGGAAT ACATCTTCTT TGGGACATGA CCCTTCTCCT 2160
 10 TCTTTTCCGT ATACATCTTT TTGGGTATTT CATGGTGATC AAACAACATT GTGATCGAAA 2220
 GCAGAGACGG CCATGGTGCT GGCTTTGAGC GTCTGGCGTT TTGTGTGTCC TGCACCTGAG 2280
 CAACCCCAAG CTGACCGCTA GGAAACTCA TTGATGTGAT TTATATCGTA CGATGAAAGA 2340
 15 GAATAAATG ATAGAAGAAC AAAGAAGAAC AAAGTAGAAG AACGTCTGAG AAGAAAGACA 2400
 GGAAATGAC ACGTACATAG TGTTCGATGA TGAATGATAT AATATTAAAT ATAAATGAG 2460
 GTAAACGTAT AGCATCACGG GATGAACGGA TGAACATGTA GTGGACAAGG TTGGGAAATA 2520
 20 GGAATGTAGA ATCCAAGAAT CGTTGACTGA TGGACGGACG TATGTAAACA GGTACACCCC 2580
 AAAGAAAAGA AAGAAAGAAA GAAAGAAAAC ACAAGCCAA GGAAGTAAAG CAGATGGTCT 2640
 TCTAAGAATA CGGCTTCAAA AAGACAGTGA ACACTCGTCG TCGAGGAATG ACAAGAAAAG 2700
 TGAGAGACTA CGAAAGGAAG AAACCAAGAC GAAAAGAAGA ACGGAGATCG AACGGACAGA 2760
 25 AATAAAG 2767

(2) INFORMATION FOR SEQ ID NO:5:

30 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 4092 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear
 35 (ii) MOLECULE TYPE: DNA (genomic)
 (iii) HYPOTHETICAL: NO
 (iv) ANTI-SENSE: NO
 (vi) ORIGINAL SOURCE:
 40 (A) ORGANISM: *Phaffia rhodozyma*
 (B) STRAIN: ATCC96594
 (ix) FEATURE:
 (A) NAME/KEY: exon
 (B) LOCATION: 852..986
 45 (ix) FEATURE:
 (A) NAME/KEY: intron
 (B) LOCATION: 987..1173
 (ix) FEATURE:
 50 (A) NAME/KEY: exon
 (B) LOCATION: 1174..1317
 (ix) FEATURE:
 (A) NAME/KEY: intron
 55

(B) LOCATION: 1318..1468

5 (ix) FEATURE:
 (A) NAME/KEY: exon
 (B) LOCATION: 1469..1549

(ix) FEATURE:
 (A) NAME/KEY: intron
 (B) LOCATION: 1550..1671

10 (ix) FEATURE:
 (A) NAME/KEY: exon
 (B) LOCATION: 1672..1794

(ix) FEATURE:
 (A) NAME/KEY: intron
 15 (B) LOCATION: 1795..1890

(ix) FEATURE:
 (A) NAME/KEY: exon
 (B) LOCATION: 1891..1979

20 (ix) FEATURE:
 (A) NAME/KEY: intron
 (B) LOCATION: 1980..2092

(ix) FEATURE:
 (A) NAME/KEY: exon
 (B) LOCATION: 2093..2165

25 (ix) FEATURE:
 (A) NAME/KEY: intron
 (B) LOCATION: 2166..2250

(ix) FEATURE:
 (A) NAME/KEY: exon
 30 (B) LOCATION: 2251..2391

(ix) FEATURE:
 (A) NAME/KEY: intron
 (B) LOCATION: 2392..2488

35 (ix) FEATURE:
 (A) NAME/KEY: exon
 (B) LOCATION: 2489..2652

(ix) FEATURE:
 (A) NAME/KEY: intron
 40 (B) LOCATION: 2653..2784

(ix) FEATURE:
 (A) NAME/KEY: exon
 (B) LOCATION: 2785..2902

45 (ix) FEATURE:
 (A) NAME/KEY: polyA_site
 (B) LOCATION: 3024

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

CGCCCGGTAT CTTGCCACAG ATGCCGCCGG AGTGCTGGC GGAGTGCTAG GAACAACGTC	60
50 ATCTCCATCT GACGAGCAAG CGTACCACAA GCTAGCTCTT CGTCTGTCAG AAGGACATCC	120
ACGCACCTTC CTGGCCTTCG GGGATGGCAC CTTCTCGTCG ACTTCCCATG GCCGTGCCCC	180

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EP 0 955 363 A2

	TGGCCTTG TG AAGATACTGT TTGCCAAGCT GAGCGCCTCC CCGCTGCTCC AGGTCCGCAA	240
5	GGTCCGAGAG TATTGGACGT CGAAGATATG TTCAAAGTGT CAGGCGAGTT CTCGGGAGAA	300
	AAAAAAGCG TGGGCTCTGA AACAGTGTGG AAATGTCTAC AAAGTGAGCT GGATTTATTG	360
	TGTGTGTATG TGTGTGTGTG TGTATGTTCT GTGTTGGTTG CTCACTGTAC TCTATGCTCT	420
	CTCTTAGATT TGGGGAACAG TGCTGTGAAC GCGTCGCGAA ACATGCTGCA CCTAGCCCTT	480
10	CACCAGAAGG AGAACCAGAG GCGGGAATG CTGGTGTCTG ACGCTGCTAC TGCTGCTACG	540
	CTAGCCGCTG AGGCTGAGGC TGGCAGAAAC TAAATCCATG ACCCATCAGA TCTTGGTGAT	600
	TCGTGGTCTG AGGACACCCA AGTCCAAAAG GGCTATATAT CGACCATCAT CCGTTGCGGT	660
15	CACTCAGTAG TAACTAAAGC TATACATAGG AATGTTCTGA ACTTGATAAC CCTAACACTA	720
	CGAAAATATC TCGGAAAATA GATTAATTTT CTTCTCATCT CAAACAAAAG ACACAACACC	780
	ATCAATCAGC CTCCTTTTAC ACACCTCTCT TTTTGTCTCT TCGTTGACAA GAAAATAACA	840
20	TCAATAGCCA AATGTCCACT ACGCCTGAAG AGAAGAAAGC AGCTCGAGCA AAGTTCGAGG	900
	CTGTCTTCCC GGTCAATGCG GATGAGATTC TCGATTATAT GAAGGGTGAA GGCATGCCCTG	960
	CCGAGGCTTT GGAATGGATG AACAAAGGTT GTCAAGGGTT TCTTCTTTAT TCTTCTGGTC	1020
25	TTTGTTCGCG TCGAACTGGC TTTCGAACTT GGCTTGACCG GGTGGGATCT CGGTGTGTGC	1080
	GCCAAAACGA TGTGGAAGCA AAACCTACTC TTACCTGTTT GGTTCCTTCT CTTCCGACCT	1140
	TCTCTCTACC CTTCGCTCCG ATCGGTCTTA TAGAAGTTGT ACTACAACAC TCCCGGAGGA	1200
30	AAACTCAACC GAGGACTTTT CGTGGTGGAT ACTTATATCC TTCTCTCGCC TTCTGGAAAA	1260
	GACATCTCGG AAGAAGAGTA CTTGAAGGCC GCTATCCTCG GTTGGTGATAT CGAGCTTGTA	1320
	CGCGTTTCTT TCATTACCTT TTCTTTCTCG TCTTCTACTC TCTTCTCTCG AACTATCTTC	1380
	CCTGCGTGTC ATCCTACAGG AATCTTTTATA CTTACATGTT GGAACATATG CCCTGTTCTT	1440
35	AATTCACCTC TTTGTCTCTG GATGGTAGCT CCAAGCTTAC TTCTTGCTGG CTGATGATAT	1500
	GATGGACGCC TCAATCACC GACGAGGCCA ACCCTGTTGG TACAAAGTTG TTAGTCCCTT	1560
	CTTCTCTTTC TGTCCCTCTT CTTCTGAGCT ATGCCAATTC TTGATTGAAA TCGGTGGTGC	1620
40	CGTCCGACT AATCCGTTTG TCGTTTTTAT CATATCTTCT TGCACAAACA GGAGGGAGTG	1680
	TCTAACATG CCATCAACGA CGCGTTCATG CTCGAGGGAG CTATCTACTT TTTGCTCAAG	1740
	AAGCACTTCC GAAAGCAGAG CTAATATGTC GATCTGCTAG AGCTCTTCCA CGATGTTTGT	1800
45	CTCTATTTCT TTTCTTCTCT CCCTCAATAA ACTGTATTTG TGACCATCTT GGATCCTTTC	1860
	CTGACGATGA ATCATTCTTC GGATGAGTAG GTTACTTTCC AAACCGAGTT GGGACAGCTC	1920
	ATCGATCTGT TGACCGCTCC TGAGGATCAC GTCGATCTCG ACAAGTTCTC CCTTAACAAG	1980
50	TATGCCCGTC ATATATTGCT TTTGTTGTCAT TCACGTCTGA TTGTCAGCTC CGATTATTGA	2040
	CTCTGATGGT GATGGTATTG ACCACATCAT GCGATGTTTG ACTTTCTCGT AGGCACCACC	2100
	TCATCGTTGT TTACAAGACC GCTTTCTATT CATTTACCTT TCCTGTGCA CTCGCTATGC	2160
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EP 0 955 363 A2

	GAATGGTGGG TCTCTCTCTT CAACTGTTCT TCCTGATTTT CTTGACCATC TGTAACATAA	2220
	ATCCTTGGA TTTTGAATC TATGTCATAG GTCGGCGTGA CAGATGAGGA GGCGTACAAG	2280
5	CTTGCGCTCT CGATCCTCAT CCCGATGGGT GAATACTTTC AAGTTCAGGA TGATGTGCTC	2340
	GACGCGTTCG CTCCTCCGGA GATCCTTGGA AAGATCGGAA CCGACATCTT GGTGCGTTTT	2400
	CGTTCCCTTC TTCTACGTTT TGTTTTCTAT CTCTGACTC CCCGTCCATC ATTTATGCTT	2460
10	CTGTTAAAC GTATTGAAAC ATCAAAAGGA CAACAAATGT TCATGGCCTA TCAACCTTGC	2520
	ACTCTCTCTC GCCTCGCCCG CTCAGCGAGA GATTCTCGAT ACTTCGTACG GTCAGAAGAA	2580
	CTCGGAGGCA GAGGCCAGAG TCAAGGCTCT GTACGCTGAG CTTGATATCC AGGGAAGTT	2640
15	CAACGCTTAT GAGTATGTCA TCTTTTAA ATTTCTAAT TTTCTTTTCA TCTCTTGTC	2700
	CCAAGAATTA TTTTGTGAAA GTTCTGGGAC TGAACATGGT GCATCCCTTT GGGTTCACCTC	2760
	CGCATATGTC TCCCGTTTGA ATAGGCAACA GAGTTACGAG TCGCTGAACA AGTTGATTGA	2820
20	CAGTATTGAC GAAGAGAAGA GTGGACTCAA GAAAGAAGTC TTCCACAGCT TCCTGGGTAA	2880
	GGTCTATAAG CGAAGCAAGT AATTCTCCTC TTATATGCA AAGGGAAGAT TTTGGCGGGA	2940
	GTGATAGGTA GGAAGAGAAG GGAGGGTCAT ATTCATTAGG CATTTCTCTT GCAGATATAG	3000
	ATGATCAAAA AGGGATATCG GTCCTCTTCT TTGTTCCGAA TACATAATAA GTCATACGAA	3060
25	GCCGAACATG AAAAAAGTGG TTCATGAGAT CAAACTTTT GCATGATCTT CTGCGATTTT	3120
	GTACAATTCT CTCGCATCCT ATTAGGATCG AACCAGGAGA AGATGAGAGA AGGAAACCCT	3180
	CACCCCGTCA GATAACAAAC GAGAAGTCTC ATCACACACA CACACAGATG AAAGAGAAAA	3240
30	ATAAAGTAC GAGGATAACT TCCAATCCGA TTTTCCAGC CCACGAACCT TCCTTGGTCC	3300
	CCGCTCCGGT GCCTTCGAGT CCGATCAATG GGGCCCAAC GCCTGAAGAT CCAAAGAACC	3360
	CTTGTTGAGG TGTATTTCTC GTCTGAGCAA TCTTAGATCC TTCAATTGTC AGTCGCGCAT	3420
35	ATATACCATC AACATCATCG TCATCACCAT CATTTGTCGTC CACAACAGCA CCGCAACGCC	3480
	GTTAATGGCA GGGCTTGAC AACTTGAGGC GGTTCCTAGC AGGTCGGACC GATTGGAGCT	3540
	CGACCCAGGG TGCACATCAC CAAGACACAT TCTCCTCAA ATGAGCGAAC AAGACATAAT	3600
40	GAGGGAAGTA GTACGCTATC GAACGTCTT TCACATCCCG GGTTCCTGGC GTATCTTTTG	3660
	GCGATTTCTT TTGTTGAAAT AGAAAATTGA AGAGAAAAAA AGAGATCCAC ATGATGAAGA	3720
	ACGGCTCTGT AGATTTCATC TCGAAAGAAA GAAAGAAAGA AAAAGAGGGG AACGAACGGA	3780
45	TCTGAATCTG TGGCCAACCA AAAAGTAGGC ACAAAGATGA CAACAGCGCC CTCTTCGACA	3840
	AGTCTTTGAA CTGCTTGTGG ATGAGACAAG TCCAGCAGA TCAACATTCC TGCTTTACCC	3900
	CATGGAGTAT CAAACACCTG AGAATAGGTC TTGCCCCTGT GTAGATAATC TCTGGACCGT	3960
	CATATGCGCG AAACGATCAG TACGACCGAC TCTACTCGAA GTCGTCAAGA GCACGGACGA	4020
50	GAACGAAAAG AGGACAAACC GCTCTGGATG CCATAAATTT CTCTTCTCAT ACCTCTCCCA	4080
	CCCACCTCA GG	4092

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 1091 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: Phaffia rhodozyma
 (B) STRAIN: ATCC96594

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

Met Tyr Thr Ile Lys His Ser Asn Phe Leu Ser Gln Thr Ile Ser Thr
 1 5 10 15

Gln Ser Thr Thr Ser Trp Val Val Asp Ala Phe Phe Ser Leu Gly Ser
 20 25 30

Arg Tyr Leu Asp Leu Ala Lys Gln Ala Asp Ser Ala Asp Ile Phe Met
 35 40 45

Val Leu Leu Gly Tyr Val Leu Met His Gly Thr Phe Val Arg Leu Phe
 50 55 60

Leu Asn Phe Arg Arg Met Gly Ala Asn Phe Trp Leu Pro Gly Met Val
 65 70 75 80

Leu Val Ser Ser Ser Phe Ala Phe Leu Thr Ala Leu Leu Ala Ala Ser
 85 90 95

Ile Leu Asn Val Pro Ile Asp Pro Ile Cys Leu Ser Glu Ala Leu Pro
 100 105 110

Phe Leu Val Leu Thr Val Gly Phe Asp Lys Asp Phe Thr Leu Ala Lys
 115 120 125

Ser Val Phe Ser Ser Pro Glu Ile Ala Pro Val Met Leu Arg Arg Lys
 130 135 140

Pro Val Ile Gln Pro Gly Asp Asp Asp Asp Leu Glu Gln Asp Glu His
 145 150 155 160

Ser Arg Val Ala Ala Asn Lys Val Asp Ile Gln Trp Ala Pro Pro Val
 165 170 175

Ala Ala Ser Arg Ile Val Ile Gly Ser Val Glu Lys Ile Gly Ser Ser
 180 185 190

Ile Val Arg Asp Phe Ala Leu Glu Val Ala Val Leu Leu Leu Gly Ala
 195 200 205

Ala Ser Gly Leu Gly Gly Leu Lys Glu Phe Cys Lys Leu Ala Ala Leu
 210 215 220

Ile Leu Val Ala Asp Cys Cys Phe Thr Phe Thr Phe Tyr Val Ala Ile
 225 230 235 240

Leu Thr Val Met Val Glu Val His Arg Ile Lys Ile Ile Arg Gly Phe
 245 250 255

EP 0 955 363 A2

Arg Pro Ala His Asn Asn Arg Thr Pro Asn Thr Val Pro Ser Thr Pro
 260 265 270
 5 Thr Ile Asp Gly Gln Ser Thr Asn Arg Ser Gly Ile Ser Ser Gly Pro
 275 280 285
 Pro Ala Arg Pro Thr Val Pro Val Trp Lys Lys Val Trp Arg Lys Leu
 290 295 300
 10 Met Gly Pro Glu Ile Asp Trp Ala Ser Glu Ala Glu Ala Arg Asn Pro
 305 310 315 320
 Val Pro Lys Leu Lys Leu Leu Leu Ile Leu Ala Phe Leu Ile Leu His
 325 330 335
 15 Ile Leu Asn Leu Cys Thr Pro Leu Thr Glu Thr Thr Ala Ile Lys Arg
 340 345 350
 Ser Ser Ser Ile His Gln Pro Ile Tyr Ala Asp Pro Ala His Pro Ile
 355 360 365
 20 Ala Gln Thr Asn Thr Thr Leu His Arg Ala His Ser Leu Val Ile Phe
 370 375 380
 Asp Gln Phe Leu Ser Asp Trp Thr Thr Ile Val Gly Asp Pro Ile Met
 385 390 395 400
 Ser Lys Trp Ile Ile Ile Thr Leu Gly Val Ser Ile Leu Leu Asn Gly
 405 410 415
 25 Phe Leu Leu Lys Gly Ile Ala Ser Gly Ser Ala Leu Gly Pro Gly Arg
 420 425 430
 Ala Gly Gly Gly Gly Ala Ala Ala Ala Ala Val Leu Leu Gly Ala
 435 440 445
 30 Trp Glu Ile Val Asp Trp Asn Asn Glu Thr Glu Thr Ser Thr Asn Thr
 450 455 460
 Pro Ala Gly Pro Pro Gly His Lys Asn Gln Asn Val Asn Leu Arg Leu
 465 470 475 480
 35 Ser Leu Glu Arg Asp Thr Gly Leu Leu Arg Tyr Gln Arg Glu Gln Ala
 485 490 495
 Tyr Gln Ala Gln Ser Gln Ile Leu Ala Pro Ile Ser Pro Val Ser Val
 500 505 510
 40 Ala Pro Val Val Ser Asn Gly Asn Gly Asn Ala Ser Lys Ser Ile Glu
 515 520 525
 Lys Pro Met Pro Arg Leu Val Val Pro Asn Gly Pro Arg Ser Leu Pro
 530 535 540
 45 Glu Ser Pro Pro Ser Thr Thr Glu Ser Thr Pro Val Asn Lys Val Ile
 545 550 555 560
 Ile Gly Gly Pro Ser Asp Arg Pro Ala Leu Asp Gly Leu Ala Asn Gly
 565 570 575
 50 Asn Gly Ala Val Pro Leu Asp Lys Gln Thr Val Leu Gly Met Arg Ser
 580 585 590
 Ile Glu Glu Cys Glu Glu Ile Met Lys Ser Gly Leu Gly Pro Tyr Ser
 595 600 605

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EP 0 955 363 A2

	Leu	Asn	Asp	Glu	Glu	Leu	Ile	Leu	Leu	Thr	Gln	Lys	Gly	Lys	Ile	Pro
	610						615					620				
5	Pro	Tyr	Ser	Leu	Glu	Lys	Ala	Leu	Gln	Asn	Cys	Glu	Arg	Ala	Val	Lys
	625					630					635					640
	Ile	Arg	Arg	Ala	Val	Ile	Ser	Arg	Ala	Ser	Val	Thr	Lys	Thr	Leu	Glu
					645					650					655	
10	Thr	Ser	Asp	Leu	Pro	Met	Lys	Asp	Tyr	Asp	Tyr	Ser	Lys	Val	Met	Gly
				660					665					670		
	Ala	Cys	Cys	Glu	Asn	Val	Val	Gly	Tyr	Met	Pro	Leu	Pro	Val	Gly	Ile
				675				680					685			
15	Ala	Gly	Pro	Leu	Asn	Ile	Asp	Gly	Glu	Val	Val	Pro	Ile	Pro	Met	Ala
	690						695					700				
	Thr	Thr	Glu	Gly	Thr	Leu	Val	Ala	Ser	Thr	Ser	Arg	Gly	Cys	Lys	Ala
	705					710					715					720
	Leu	Asn	Ala	Gly	Gly	Gly	Val	Thr	Thr	Val	Ile	Thr	Gln	Asp	Ala	Met
					725					730					735	
20	Thr	Arg	Gly	Pro	Val	Val	Asp	Phe	Pro	Ser	Val	Ser	Gln	Ala	Ala	Gln
				740					745					750		
	Ala	Lys	Arg	Trp	Leu	Asp	Ser	Val	Glu	Gly	Met	Glu	Val	Met	Ala	Ala
				755				760					765			
25	Ser	Phe	Asn	Ser	Thr	Ser	Arg	Phe	Ala	Arg	Leu	Gln	Ser	Ile	Lys	Cys
	770						775					780				
	Gly	Met	Ala	Gly	Arg	Ser	Leu	Tyr	Ile	Arg	Leu	Ala	Thr	Ser	Thr	Gly
	785					790				795						800
30	Asp	Ala	Met	Gly	Met	Asn	Met	Ala	Gly	Lys	Gly	Thr	Glu	Lys	Ala	Leu
					805					810					815	
	Glu	Thr	Leu	Ser	Glu	Tyr	Phe	Pro	Ser	Met	Gln	Ile	Leu	Ala	Leu	Ser
				820					825					830		
35	Gly	Asn	Tyr	Cys	Ile	Asp	Lys	Lys	Pro	Ser	Ala	Ile	Asn	Trp	Ile	Glu
			835				840						845			
	Gly	Arg	Gly	Lys	Ser	Val	Val	Ala	Glu	Ser	Val	Ile	Pro	Gly	Ala	Ile
		850					855					860				
40	Val	Lys	Ser	Val	Leu	Lys	Thr	Thr	Val	Ala	Asp	Leu	Val	Asn	Leu	Asn
	865					870					875					880
	Ile	Lys	Lys	Asn	Leu	Ile	Gly	Ser	Ala	Met	Ala	Gly	Ser	Ile	Gly	Gly
				885						890					895	
45	Phe	Asn	Ala	His	Ala	Ser	Asp	Ile	Leu	Thr	Ser	Ile	Phe	Leu	Ala	Thr
				900					905					910		
	Gly	Gln	Asp	Pro	Ala	Gln	Asn	Val	Glu	Ser	Ser	Met	Cys	Met	Thr	Leu
			915					920					925			
	Met	Glu	Ala	Val	Asn	Asp	Gly	Lys	Asp	Leu	Leu	Ile	Thr	Cys	Ser	Met
		930					935					940				
50	Pro	Ala	Ile	Glu	Cys	Gly	Thr	Val	Gly	Gly	Gly	Thr	Phe	Leu	Pro	Pro
	945					950					955					960

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(2) INFORMATION FOR SEQ ID NO:7:

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

EP 0 955 363 A2

	Asn Thr Asp Ile Glu Gly Ile Asp Ser Lys Asn Ala Cys Tyr Gly Ser	115	120	125
5	Thr Ala Ala Leu Phe Asn Ala Val Asn Trp Ile Glu Ser Ser Ser Trp	130	135	140
	Asp Gly Arg Asn Ala Ile Val Phe Cys Gly Asp Ile Ala Ile Tyr Ala	145	150	155
10	Glu Gly Ala Ala Arg Pro Ala Gly Gly Ala Gly Ala Cys Ala Ile Leu	165	170	175
	Ile Gly Pro Asp Ala Pro Val Val Phe Glu Pro Val His Gly Asn Phe	180	185	190
15	Met Thr Asn Ala Trp Asp Phe Tyr Lys Pro Asn Leu Ser Ser Glu Tyr	195	200	205
	Pro Ile Val Asp Gly Pro Leu Ser Val Thr Ser Tyr Val Asn Ala Ile	210	215	220
20	Asp Lys Ala Tyr Glu Ala Tyr Arg Thr Lys Tyr Ala Lys Arg Phe Gly	225	230	235
	Gly Pro Lys Thr Asn Gly Val Thr Asn Gly His Thr Glu Val Ala Gly	245	250	255
25	Val Ser Ala Ala Ser Phe Asp Tyr Leu Leu Phe His Ser Pro Tyr Gly	260	265	270
	Lys Gln Val Val Lys Gly His Gly Arg Leu Leu Tyr Asn Asp Phe Arg	275	280	285
30	Asn Asn Pro Asn Asp Pro Val Phe Ala Glu Val Pro Ala Glu Leu Ala	290	295	300
	Thr Leu Asp Met Lys Lys Ser Leu Ser Asp Lys Asn Val Glu Lys Ser	305	310	315
35	Leu Ile Ala Ala Ser Lys Ser Ser Phe Asn Lys Gln Val Glu Pro Gly	325	330	335
	Met Thr Thr Val Arg Gln Leu Gly Asn Leu Tyr Thr Ala Ser Leu Phe	340	345	350
40	Gly Ala Leu Ala Ser Leu Phe Ser Asn Val Pro Gly Asp Glu Leu Val	355	360	365
	Gly Lys Arg Ile Ala Leu Tyr Ala Tyr Gly Ser Gly Ala Ala Ala Ser	370	375	380
45	Phe Tyr Ala Leu Lys Val Lys Ser Ser Thr Ala Phe Ile Ser Glu Lys	385	390	395
	Leu Asp Leu Asn Asn Arg Leu Ser Asn Met Lys Ile Val Pro Cys Asp	405	410	415
50	Asp Phe Val Lys Ala Leu Lys Val Arg Glu Glu Thr His Asn Ala Val	420	425	430
	Ser Tyr Ser Pro Ile Gly Ser Leu Asp Asp Leu Trp Pro Gly Ser Tyr	435	440	445
55	Tyr Leu Gly Glu Ile Asp Ser Met Trp Arg Arg Gln Tyr Lys Gln Val	450	455	460

Pro Ser Ala
465

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 432 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:
 (A) ORGANISM: *Phaffia rhodozyma*
 (B) STRAIN: ATCC96594

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

Lys	Glu	Glu	Ile	Leu	Val	Ser	Ala	Pro	Gly	Lys	Val	Ile	Leu	Phe	Gly	1	5	10	15
Glu	His	Ala	Val	Gly	His	Gly	Val	Thr	Gly	Ile	Ala	Ala	Ser	Val	Asp	20	25	30	
Leu	Arg	Cys	Tyr	Ala	Leu	Leu	Ser	Pro	Thr	Ala	Thr	Thr	Thr	Thr	Ser	35	40	45	
Ser	Ser	Leu	Ser	Ser	Thr	Asn	Ile	Thr	Ile	Ser	Leu	Thr	Asp	Leu	Asn	50	55	60	
Phe	Thr	Gln	Ser	Trp	Pro	Val	Asp	Ser	Leu	Pro	Trp	Ser	Leu	Ala	Pro	65	70	75	80
Asp	Trp	Thr	Glu	Ala	Ser	Ile	Pro	Glu	Ser	Leu	Cys	Pro	Thr	Leu	Leu	85	90	95	
Ala	Glu	Ile	Glu	Arg	Ile	Ala	Gly	Gln	Gly	Gly	Asn	Gly	Gly	Glu	Arg	100	105	110	
Glu	Lys	Val	Ala	Thr	Met	Ala	Phe	Leu	Tyr	Leu	Leu	Val	Leu	Leu	Ser	115	120	125	
Lys	Gly	Lys	Pro	Ser	Glu	Pro	Phe	Glu	Leu	Thr	Ala	Arg	Ser	Ala	Leu	130	135	140	
Pro	Met	Gly	Ala	Gly	Leu	Gly	Ser	Ser	Ala	Ala	Leu	Ser	Thr	Ser	Leu	145	150	155	160
Ala	Leu	Val	Phe	Leu	Leu	His	Phe	Ser	His	Leu	Ser	Pro	Thr	Thr	Thr	165	170	175	
Gly	Arg	Glu	Ser	Thr	Ile	Pro	Thr	Ala	Asp	Thr	Glu	Val	Ile	Asp	Lys	180	185	190	
Trp	Ala	Phe	Leu	Ala	Glu	Lys	Val	Ile	His	Gly	Asn	Pro	Ser	Gly	Ile	195	200	205	
Asp	Asn	Ala	Val	Ser	Thr	Arg	Gly	Gly	Ala	Val	Ala	Phe	Lys	Arg	Lys	210	215	220	
Ile	Glu	Gly	Lys	Gln	Glu	Gly	Gly	Met	Glu	Ala	Ile	Lys	Ser	Phe	Thr	225	230	235	240

EP 0 955 363 A2

Ser Ile Arg Phe Leu Ile Thr Asp Ser Arg Ile Gly Arg Asp Thr Arg
 245 250 255
 5 Ser Leu Val Ala Gly Val Asn Ala Arg Leu Ile Gln Glu Pro Glu Val
 260 265 270
 Ile Val Pro Leu Leu Glu Ala Ile Gln Gln Ile Ala Asp Glu Ala Ile
 275 280 285
 10 Arg Cys Leu Lys Asp Ser Glu Met Glu Arg Ala Val Met Ile Asp Arg
 290 295 300
 Leu Gln Asn Leu Val Ser Glu Asn His Ala His Leu Ala Ala Leu Gly
 305 310 315 320
 15 Val Ser His Pro Ser Leu Glu Glu Ile Ile Arg Ile Gly Ala Asp Lys
 325 330 335
 Pro Phe Glu Leu Arg Thr Lys Leu Thr Gly Ala Gly Gly Gly Cys
 340 345 350
 20 Ala Val Thr Leu Val Pro Asp Asp Phe Ser Thr Glu Thr Leu Gln Ala
 355 360 365
 Leu Met Glu Thr Leu Val Gln Ser Ser Phe Ala Pro Tyr Ile Ala Arg
 370 375 380
 Val Gly Gly Ser Gly Val Gly Phe Leu Ser Ser Thr Lys Ala Asp Pro
 385 390 395 400
 25 Glu Asp Gly Glu Asn Arg Leu Lys Asp Gly Leu Val Gly Thr Glu Ile
 405 410 415
 Asp Glu Leu Asp Arg Trp Ala Leu Lys Thr Gly Arg Trp Ser Phe Ala
 420 425 430

30

(2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 401 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear
 35
 (ii) MOLECULE TYPE: protein
 (iii) HYPOTHETICAL: NO
 40 (vi) ORIGINAL SOURCE:
 (A) ORGANISM: Phaffia rhodozyma
 (B) STRAIN: ATCC96594

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Met Val His Ile Ala Thr Ala Ser Ala Pro Val Asn Ile Ala Cys Ile
 1 5 10 15
 45 Lys Tyr Trp Gly Lys Arg Asp Thr Lys Leu Ile Leu Pro Thr Asn Ser
 20 25 30
 Ser Leu Ser Val Thr Leu Asp Gln Asp His Leu Arg Ser Thr Thr Ser
 35 40 45
 50 Ser Ala Cys Asp Ala Ser Phe Glu Lys Asp Arg Leu Trp Leu Asn Gly
 50 55 60

55

EP 0 955 363 A2

Ile Glu Glu Glu Val Lys Ala Gly Gly Arg Leu Asp Val Cys Ile Lys
 65 70 75 80
 5 Glu Met Lys Lys Leu Arg Ala Gln Glu Glu Glu Lys Asp Ala Gly Leu
 85 90 95
 Glu Lys Leu Ser Ser Phe Asn Val His Leu Ala Ser Tyr Asn Asn Phe
 100 105 110
 10 Pro Thr Ala Ala Gly Leu Ala Ser Ser Ala Ser Gly Leu Ala Ala Leu
 115 120 125
 Val Ala Ser Leu Ala Ser Leu Tyr Asn Leu Pro Thr Asn Ala Ser Glu
 130 135 140
 15 Leu Ser Leu Ile Ala Arg Gln Gly Ser Gly Ser Ala Cys Arg Ser Leu
 145 150 155 160
 Phe Gly Gly Phe Val Ala Trp Glu Gln Gly Lys Leu Ser Ser Gly Thr
 165 170 175
 20 Asp Ser Phe Ala Val Gln Val Glu Pro Arg Glu His Trp Pro Ser Leu
 180 185 190
 His Ala Leu Ile Cys Val Val Ser Asp Glu Lys Lys Thr Thr Ala Ser
 195 200 205
 25 Thr Ala Gly Met Gln Thr Thr Val Asn Thr Ser Pro Leu Leu Gln His
 210 215 220
 Arg Ile Glu His Val Val Pro Ala Arg Met Glu Ala Ile Thr Gln Ala
 225 230 235 240
 Ile Arg Ala Lys Asp Phe Asp Ser Phe Ala Lys Ile Thr Met Lys Asp
 245 250 255
 30 Ser Asn Gln Phe His Ala Val Cys Leu Asp Ser Glu Pro Pro Ile Phe
 260 265 270
 Tyr Leu Asn Asp Val Ser Arg Ser Ile Ile His Leu Val Thr Glu Leu
 275 280 285
 35 Asn Arg Val Ser Val Gln Ala Gly Gly Pro Val Leu Ala Ala Tyr Thr
 290 295 300
 Phe Asp Ala Gly Pro Asn Ala Val Ile Tyr Ala Glu Glu Ser Ser Met
 305 310 315 320
 40 Pro Glu Ile Ile Arg Leu Ile Glu Arg Tyr Phe Pro Leu Gly Thr Ala
 325 330 335
 Phe Glu Asn Pro Phe Gly Val Asn Thr Glu Gly Gly Asp Ala Leu Arg
 340 345 350
 45 Glu Gly Phe Asn Gln Asn Val Ala Pro Val Phe Arg Lys Gly Ser Val
 355 360 365
 Ala Arg Leu Ile His Thr Arg Ile Gly Asp Gly Pro Arg Thr Tyr Gly
 370 375 380
 50 Glu Glu Glu Ser Leu Ile Gly Glu Asp Gly Leu Pro Lys Val Val Lys
 385 390 395 400
 Ala

55

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 355 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:

- (A) ORGANISM: *Phaffia rhodozyma*
 (B) STRAIN: ATCC96594

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

```

Met Ser Thr Thr Pro Glu Glu Lys Lys Ala Ala Arg Ala Lys Phe Glu
1          5          10          15
Ala Val Phe Pro Val Ile Ala Asp Glu Ile Leu Asp Tyr Met Lys Gly
20          25          30
Glu Gly Met Pro Ala Glu Ala Leu Glu Trp Met Asn Lys Asn Leu Tyr
35          40          45
Tyr Asn Thr Pro Gly Gly Lys Leu Asn Arg Gly Leu Ser Val Val Asp
50          55          60
Thr Tyr Ile Leu Leu Ser Pro Ser Gly Lys Asp Ile Ser Glu Glu Glu
65          70          75          80
Tyr Leu Lys Ala Ala Ile Leu Gly Trp Cys Ile Glu Leu Leu Gln Ala
85          90          95
Tyr Phe Leu Val Ala Asp Asp Met Met Asp Ala Ser Ile Thr Arg Arg
100          105          110
Gly Gln Pro Cys Trp Tyr Lys Val Glu Gly Val Ser Asn Ile Ala Ile
115          120          125
Asn Asn Ala Phe Met Leu Glu Gly Ala Ile Tyr Phe Leu Leu Lys Lys
130          135          140
His Phe Arg Lys Gln Ser Tyr Tyr Val Asp Leu Leu Glu Leu Phe His
145          150          155          160
Asp Val Thr Phe Gln Thr Glu Leu Gly Gln Leu Ile Asp Leu Leu Thr
165          170          175
Ala Pro Glu Asp His Val Asp Leu Asp Lys Phe Ser Leu Asn Lys His
180          185          190
His Leu Ile Val Val Tyr Lys Thr Ala Phe Tyr Ser Phe Tyr Leu Pro
195          200          205
Val Ala Leu Ala Met Arg Met Val Gly Val Thr Asp Glu Glu Ala Tyr
210          215          220
Lys Leu Ala Leu Ser Ile Leu Ile Pro Met Gly Glu Tyr Phe Gln Val
225          230          235          240
Gln Asp Asp Val Leu Asp Ala Phe Arg Pro Pro Glu Ile Leu Gly Lys
245          250          255

```


EP 0 955 363 A2

Ile Gly Thr Asp Ile Leu Asp Asn Lys Cys Ser Trp Pro Ile Asn Leu
260 265 270
5 Ala Leu Ser Pro Ala Ser Pro Ala Gln Arg Glu Ile Leu Asp Thr Ser
275 280 285
Tyr Gly Gln Lys Asn Ser Glu Ala Glu Ala Arg Val Lys Ala Leu Tyr
290 295 300
10 Ala Glu Leu Asp Ile Gln Gly Lys Phe Asn Ala Tyr Glu Gln Gln Ser
305 310 315 320
Tyr Glu Ser Leu Asn Lys Leu Ile Asp Ser Ile Asp Glu Glu Lys Ser
325 330 335
15 Gly Leu Lys Lys Glu Val Phe His Ser Phe Leu Gly Lys Val Tyr Lys
340 345 350
Arg Ser Lys
355

(2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 26 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

GGNAARTAYA CNATHGGNYT NGGNCA

26

(2) INFORMATION FOR SEQ ID NO:12:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 26 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

TANARNSWNS WNGTRTACAT RTTNCC

26

(2) INFORMATION FOR SEQ ID NO:13:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 24 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

5 (iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

10 GAAGAACCCC ATCAAAAGCC TCGA 24

(2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS:

15 (A) LENGTH: 25 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

20 (iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

25 AAAAGCCTCG AGATCCTTGT GAGCG 25

(2) INFORMATION FOR SEQ ID NO:15:

(i) SEQUENCE CHARACTERISTICS:

30 (A) LENGTH: 18 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

35 (iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

40 AGAAGCCAGA AGAGAAAA 18

(2) INFORMATION FOR SEQ ID NO:16:

(i) SEQUENCE CHARACTERISTICS:

45 (A) LENGTH: 18 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

50 (iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

55

TCGTCGAGGA AAGTAGAT

18

(2) INFORMATION FOR SEQ ID NO:17:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 30 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

GGTACCATAT GTATCCTTCT ACTACCGAAC

30

(2) INFORMATION FOR SEQ ID NO:18:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 30 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

GCATGCGGAT CCTCAAGCAG AAGGGACCTG

30

(2) INFORMATION FOR SEQ ID NO:19:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 32 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

GCNTGYTCYG ARAAYGTNAT HGGNTAYATG CC

32

(2) INFORMATION FOR SEQ ID NO:20:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 32 base pairs
 - (B) TYPE: nucleic acid

(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

5 (ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

10 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:
ATCCARTDA TNGCNGCNGG YTTYTTTRTCN GT 32

(2) INFORMATION FOR SEQ ID NO:21:

15 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 25 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

20 (ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:
25 GGCCATTCCA CACTTGATGC TCTGC 25

(2) INFORMATION FOR SEQ ID NO:22:

30 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 21 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

35 (ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:
40 GGCCGATATC TTTATGGTCC T 21

(2) INFORMATION FOR SEQ ID NO:23:

45 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 26 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

50 (ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

55

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

GGTACCGAAG AAATTATGAA GAGTGG

26

(2) INFORMATION FOR SEQ ID NO:24:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 26 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

CTGCAGTCAG GCATCCACGT TCACAC

26

(2) INFORMATION FOR SEQ ID NO:25:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 29 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

GCNCCNGGNA ARGTNATHYT NTTYGGNGA

29

(2) INFORMATION FOR SEQ ID NO:26:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 29 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

CCCCANGTNS WNACNGCRTT RTCNACNCC

29

(2) INFORMATION FOR SEQ ID NO:27:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 17 base pairs

(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

5 (ii) MOLECULE TYPE: DNA (genomic)
(iii) HYPOTHETICAL: NO
(iv) ANTI-SENSE: NO
10 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

ACATGCTGTA GTCCATG

17

(2) INFORMATION FOR SEQ ID NO:28:

15 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 16 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

20 (ii) MOLECULE TYPE: DNA (genomic)
(iii) HYPOTHETICAL: NO
(iv) ANTI-SENSE: NO
25 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

ACTCGGATTC CATGGA

16

(2) INFORMATION FOR SEQ ID NO:29:

30 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 25 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

35 (ii) MOLECULE TYPE: DNA (genomic)
(iii) HYPOTHETICAL: NO
(vi) ORIGINAL SOURCE:
(A) ORGANISM: Phaffia rhodozyma
(B) STRAIN: ATCC96594
40

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

TTGTTGTCGT AGCAGTGGGT GAGAG

25

45 (2) INFORMATION FOR SEQ ID NO:30:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 18 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
50 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)
(iii) HYPOTHETICAL: NO

55

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

GGAAGAGGAA GAGAAAAG

18

(2) INFORMATION FOR SEQ ID NO:31:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 18 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:

TTGCCGAAC CAATGTAG

18

(2) INFORMATION FOR SEQ ID NO:32:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 26 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:
 (A) ORGANISM: Phaffia rhodozyma
 (B) STRAIN: ATCC96594

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:32:

GGATCCATGA GAGCCCAAAA AGAAGA

26

(2) INFORMATION FOR SEQ ID NO:33:

(i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 26 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(vi) ORIGINAL SOURCE:
 (A) ORGANISM: Phaffia rhodozyma
 (B) STRAIN: ATCC96594

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:

GTCGACTCAA GCAAAAGACC AACGAC

26

(2) INFORMATION FOR SEQ ID NO:34:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 23 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:34:

HTNAARTAYT TGGGNAARMG NGA

23

(2) INFORMATION FOR SEQ ID NO:35:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 29 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:35:

GCRTTGGNC CNGCRTCRAA NGTRTANGC

29

(2) INFORMATION FOR SEQ ID NO:36:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 20 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:36:

CCGAACTCTC GCTCATCGCC

20

(2) INFORMATION FOR SEQ ID NO:37:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 20 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:

CAGATCAGCG CGTGGAGTGA

(2) INFORMATION FOR SEQ ID NO:38:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 26 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:

CARGCNTAYT TYYTNGTNGC NGAYGA

(2) INFORMATION FOR SEQ ID NO:39:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 32 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:39:

CAYTTRTTRT CYTGDAITC NGTNCCDATY TT

(2) INFORMATION FOR SEQ ID NO:40:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 25 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:40:

ATCCTCATCC CGATGGGTGA ATACT

25

5 (2) INFORMATION FOR SEQ ID NO:41:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 25 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

10

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

15

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:41:

AGGAGCGGTC AACAGATCGA TGAGC

25

20

(2) INFORMATION FOR SEQ ID NO:42:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 25 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

25

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

30

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:42:

GAATTCATAT GTCCACTACG CCTGA

25

35

(2) INFORMATION FOR SEQ ID NO:43:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 25 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: single
 - (D) TOPOLOGY: linear

40

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

45

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:43:

GTCGACGGTA CCTATCACTC CCGCC

25

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Claims

1. An isolated DNA sequence, which codes for an enzyme involved in the mevalonate pathway or the pathway from isopentenyl pyrophosphate to farnesyl pyrophosphate.
2. An isolated DNA sequence according to claim 1, wherein said enzyme has an activity selected from the group consisting of 3-hydroxy-3-methylglutaryl-CoA synthase activity, 3-hydroxy-3-methylglutaryl-CoA reductase activity, mevalonate kinase activity, mevalonate pyrophosphate decarboxylase activity and farnesyl pyrophosphate synthase activity.
3. An isolated DNA sequence according to claim 1 or 2, which is characterized in that
 - (a) the said DNA sequence codes for the said enzyme having an amino acid sequence selected from the group consisting of those described in SEQ ID NOs: 6, 7, 8, 9 and 10, or
 - (b) the said DNA sequence codes for a variant of the said enzyme selected from (i) an allelic variant, and (ii) an enzyme having one or more amino acid addition, insertion, deletion and/or substitution and having the stated enzyme activity.
4. An isolated DNA sequence according to any one of claims 1-3, which can be derived from a gene of *Phaffia rhodozyma* and is selected from:
 - (i) a DNA sequence represented in SEQ ID NOs: 1, 2, 4 or 5;
 - (ii) an isocoding or an allelic variant for the DNA sequence represented in SEQ ID NOs: 1, 2, 4 or 5; and
 - (iii) a derivative of a DNA sequence represented in SEQ ID NOs: 1, 2, 4 or 5 with addition, insertion, deletion and/or substitution of one or more nucleotide(s), and coding for a polypeptide having the said enzyme activity.
5. An isolated DNA sequence, which is selected from:
 - (i) a DNA sequence represented in SEQ ID NO: 3;
 - (ii) an isocoding or an allelic variant for the DNA sequence represented in SEQ ID NO: 3; and
 - (iii) a derivative of a DNA sequence represented in SEQ ID NO: 3 with addition, insertion, deletion and/or substitution of one or more nucleotide(s), and coding for a polypeptide having the mevalonate kinase activity.
6. An isolated DNA sequence as claimed in claim 1 or 2 and which is selected from:
 - (i) a DNA sequence which hybridizes under standard conditions with a sequence as shown in SEQ ID Nos: 1 - 10 or its complementary strand or fragments thereof; and
 - (ii) a DNA sequence which do not hybridize as defined in (i) because of the degeneration of the genetic code but which codes for polypeptides having exactly the same amino acid sequence as shown in SEQ ID Nos: 1 - 10 or those encoded by a DNA sequence as defined above under (i).
7. A vector or plasmid comprising a DNA sequence as defined in any of claims 1-6.
8. A host cell which has been transformed or transfected by a DNA sequence as claimed in anyone of claims 1 to 6, or a vector or plasmid as claimed in claim 7.
9. A process for producing an enzyme involved in the mevalonate pathway or the pathway from isopentenyl pyrophosphate to farnesyl pyrophosphate, which comprises culturing a host cell as claimed in claim 8, under the conditions conducive to the production of said enzyme.
10. A process for the production of isoprenoids or carotenoids, preferably astaxanthin, which comprises cultivating a host cell as claimed in claim 8 under suitable culture conditions.

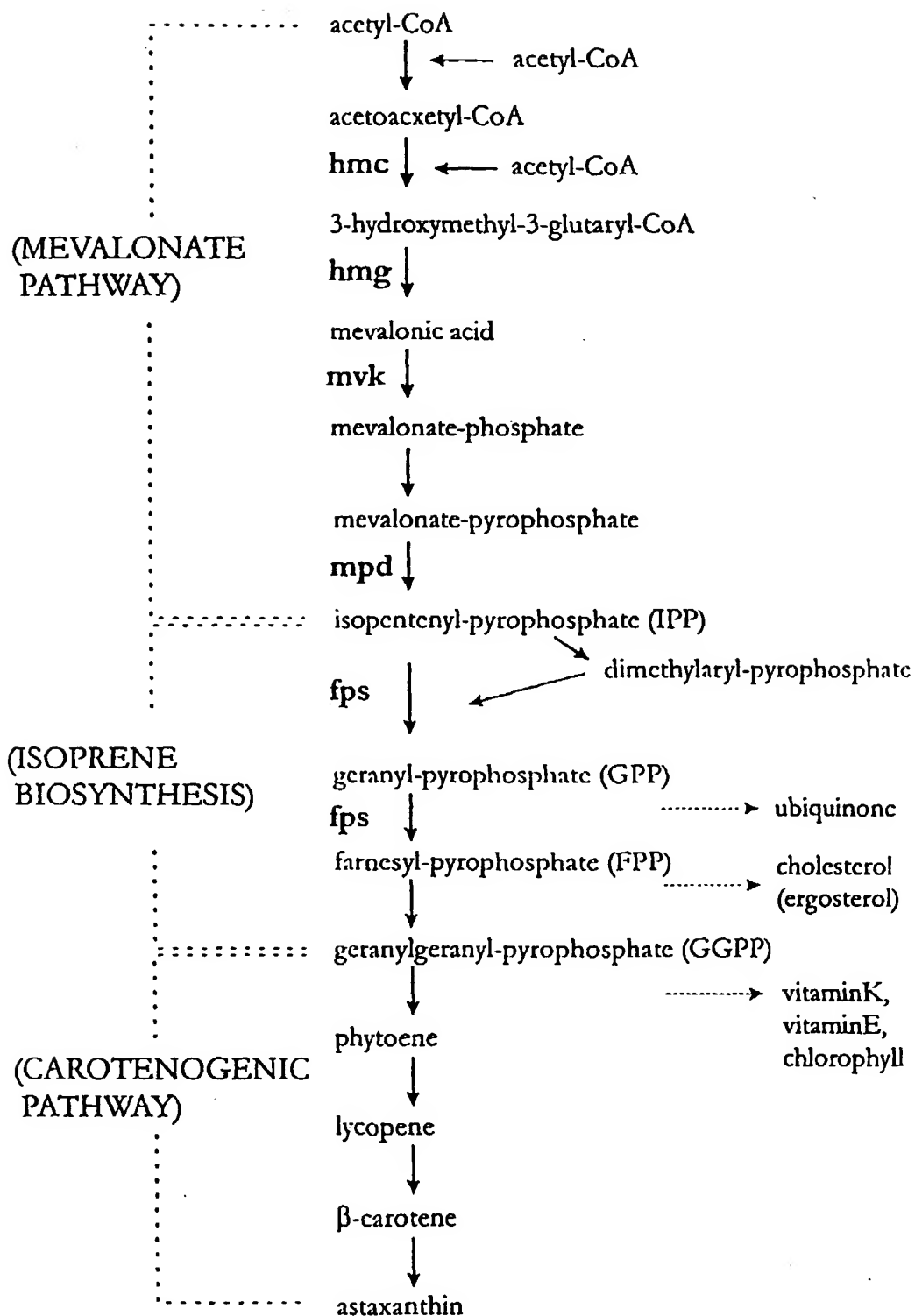
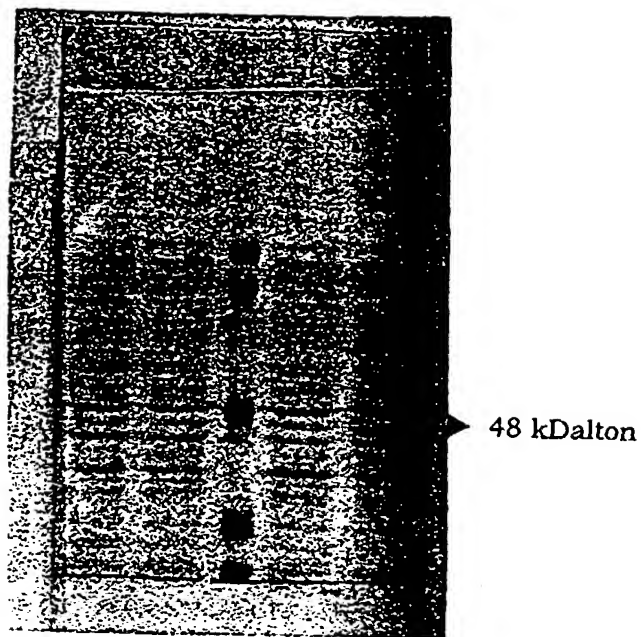


Fig. 1 Biosynthetic pathway from acetyl-CoA to astaxanthin in *P. rhodozyma*



Lane : 1 2 3 4 5

Fig. 2 Expression study of pseudo-mvk gene
by insertion of one base